Investigation of a possible outbreak of carbapenem-resistant Acinetobacter baumannii in Odense, Denmark using PFGE, MLST and whole-genome-based SNPs

Hammerum, Anette M.; Hansen, Frank; Skov, Marianne N.; Stegger, Marc; Andersen, Paal Skytt; Holm, Anette; Jakobsen, Lotte; Justesen, Ulrik Stenz

Published in:
Journal of Antimicrobial Chemotherapy

DOI:
10.1093/jac/dkv072

Publication date:
2015

Document Version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
Investigation of a possible outbreak of carbapenem-resistant 
*Acinetobacter baumannii* in Odense, Denmark using PFGE, MLST 
and whole-genome-based SNPs

Anette M. Hammerum1*, Frank Hansen1, Marianne N. Skov2, Marc Stegger1,3, Paal S. Andersen1,3,4, Anette Holm2, Lotte Jakobsen1 and Ulrik S. Justesen2

1Statens Serum Institut, Copenhagen S, Denmark; 2Department of Clinical Microbiology, Odense University Hospital, Odense C, Denmark; 3Pathogen Genomics Division, Translational Genomics Research Institute, Flagstaff, AZ, USA; 4Veterinary Disease Biology, University of Copenhagen, Copenhagen, Denmark

*Corresponding author. Department of Microbiology and Infection Control, Statens Serum Institut, Artillerivej 5 (47/219), DK-2300 Copenhagen S, Denmark. Tel: +45-3268-3399; Fax: +45-3268-3231; E-mail: ama@ssi.dk

Received 16 December 2014; returned 1 February 2015; revised 16 February 2015; accepted 25 February 2015

**Objectives:** The objectives were to study a possible outbreak of carbapenem-resistant *Acinetobacter baumannii* by comparing three different typing methods (PFGE, MLST and whole-genome SNPs) and to compare the resistance gene profiles of the isolates.

**Methods:** From December 2012 to October 2013, eight carbapenem-resistant *A. baumannii* were detected at Odense University Hospital, Odense, Denmark. These isolates were typed by PFGE, with ApaI and SmaI, respectively, and subjected to WGS. The WGS data were used for *in silico* extraction of MLST types using two different schemes, resistance genes and SNPs, to which 31 publicly available *A. baumannii* genomes were added.

**Results:** Using ApaI, the eight isolates had four different PFGE profiles, which were further differentiated using SmaI, separating one of the profiles into two distinct PFGE types. Five ST2 (Pasteur MLST) OXA-23-producing isolates, two ST1 OXA-72-producing isolates and one ST158 OXA-23-producing isolate were detected. The five ST2 isolates were subdivided into ST195, ST208 and ST218 using the Oxford MLST scheme. The phylogenetic analysis based on the core genome showed that six of the eight Danish *A. baumannii* isolates were located in three distinct clusters. The two remaining isolates did not cluster with other Danish or international isolates included in the study. Isolates that clustered using PFGE, Oxford MLST and phylogenetic analysis also shared similar resistance gene profiles.

**Conclusions:** The SNP profile, Oxford MLST, PFGE and resistance gene profiles clearly indicated spread of three different *A. baumannii* strains.

**Keywords:** antimicrobial resistance, antimicrobial resistance epidemiology, carbapenems

**Introduction**

Carbapenem resistance in *Acinetobacter baumannii* is most often encoded by oxacillinases (OXAs) and less frequently by metallob-β-lactamases.1 Four main OXA groups, which can be further subgrouped, have been associated with carbapenem resistance in *A. baumannii:* the chromosomally located intrinsic OXA-51-like group and the three acquired OXA groups—OXA-23-like, OXA-40-like and OXA-58-like.2,3 Furthermore, two novel OXA enzymes, OXA-143 and OXA-235, were detected in *A. baumannii* isolates in 2009 and 2013, respectively.4,5

Spread of *A. baumannii* in hospitals has been investigated by several typing methods. The most commonly used method for investigation of clonality is PFGE using ApaI as the restriction enzyme.5 For investigation of population structure and global bacterial epidemiology, MLST has been the golden standard.6 There are two MLST schemes for *A. baumannii:* the Oxford MLST scheme and the Pasteur MLST scheme (http://pubmlst.org/abaumannii). Furthermore, WGS can be used for comparison of *A. baumannii* isolates.

In recent years, Danish departments of clinical microbiology have, on a voluntary basis, submitted carbapenem-resistant *A. baumannii* isolates for verification and genotyping at the Antimicrobial Resistance Reference Laboratory at Statens Serum Institut.

To our knowledge, spread of OXA-23-producing *A. baumannii* has previously only been detected at one hospital in the Capital

© The Author 2015. Published by Oxford University Press on behalf of the British Society for Antimicrobial Chemotherapy. All rights reserved. For Permissions, please e-mail: journals.permissions@oup.com
Region in Denmark, whereas single cases of OXA-23-like or OXA-60-like-producing A. baumannii, most often related to prior travel, have been reported in other parts of Denmark too.6,7

The aim of this study was: (i) to study possible outbreak(s) of carbapenem-resistant A. baumannii detected at Odense University Hospital using three different typing methods [PFGE, MLST (two different schemes) and SNP analysis]; (ii) to compare these typing methods; and (iii) to compare the resistance gene profiles detected from the whole-genome sequences.

Methods

Isolates

In October 2013, carbapenem-resistant A. baumannii were detected in four patients hospitalized at Odense University Hospital, a tertiary referral hospital with approximately 1100 beds in the Region of Southern Denmark (1.2 million inhabitants).

To investigate a possible outbreak, the isolates were characterized using different typing methods. Their profiles were compared with four other carbapenem-resistant A. baumannii isolates previously detected at the hospital (December 2012 to September 2013) (Table 1). The eight isolates were the only carbapenem-resistant A. baumannii isolates detected during December 2012 to October 2013 at the Department of Clinical Microbiology at Odense University Hospital. Isolates were identified to species level by MALDI-TOF MS.

Susceptibility testing

The eight isolates were tested using Sensititre Trek panels (Thermo Scientific, Waltham, MA, USA; ampicillin/subbactam, ticarcillin/clavulanic acid, piperacillin/tazobactam, ceftazidime, cefoperazone, ceftriaxone, cefotaxime, cefepime, aztreonam, imipenem, meropenem, lomefloxacin, ciprofloxacin, levofloxacin, trimethoprim/sulfamethoxazole, gentamicin, tobramycin, amikacin, tetracycline and colistin) as per the manufacturer’s instructions. The microbroth dilution method was performed in accordance with the CLSI.8

PCR

The blaOXA genogroups in the eight carbapenem-resistant A. baumannii isolates were identified by multiplex PCR as previously described.5,10

PFGE

Bacterial DNA for PFGE was prepared using the Salmonella standard protocol of CDC PulseNet (http://www.cdc.gov/pulsenet/). Agarose plugs containing DNA were digested with 25 U of the restriction enzyme Apal or with 30 U of the restriction enzyme Smal for 4 h. Electrophoresis (1% gel) was performed at 6 V/cm and with the following run parameters for the two different enzymes: Apal, 5–20 s for 18.5 h; and Smal, 5–20 s for 18.5 h followed by 5–10 s for 4 h. Addition of thiourea (100 L of 1 M) in the running buffer was not able to stop the degradation of the DNA from two of the strains when digested with Smal (isolate AMA 517 and isolate AMA 525). The CDC standard H9812 isolate was digested with 50 U of XbaI and used as the molecular size marker. All visible bands more than 55 kb in size were included in the interpretation of PFGE patterns. An isolate was considered to be closely related to the outbreak strain if its PFGE pattern differed from the outbreak pattern by changes consistent with a single genetic event, i.e. a point mutation or any major insertions or deletions. Such changes typically result in two or three band differences. Patterns that were closely or possibly related to the outbreak pattern were considered subtypes of A and were designated type A1, type A2 etc.11

WGS and assembly

Genomic DNA was extracted (DNeasy Blood and Tissue Kit, Qiagen, Copenhagen, Denmark) and fragment libraries constructed using the Nextera Kit (Illumina, Little Chesterford, UK) followed by 251 bp paired-end sequencing (MiSeq, Illumina) according to the manufacturer’s instructions.

In addition to the eight genomes of the Danish A. baumannii isolates (NCBI BioProject ID PRJNA266271), the chromosomes of the 31 publicly available complete and scaffold A. baumannii genomes, representing 31 different genome groups according to GenBank, were included in this study. The sequence data were aligned against the chromosome of the A. baumannii BJAB0715 reference genome (GenBank accession ID CP003847) using the Burrows–Wheeler Aligner (BWA). Identification of SNP variants was performed using the GATK Unified Genotyper with filtering using NASP (http://tgennorth.github.io/NASP/) to remove positions with less than 10x coverage and <90% unambiguous variant calls, or within duplicated regions of the reference using NUCmer. Phylogenetic analyses of the identified SNPs was performed using MEGA 6.0.6.12 The paired-end Illumina data were assembled using CLCbio’s Genomic Workbench 7.5 (Qiagen, Aarhus, Denmark).

MLST

MLST was performed from the whole-genome sequences of the eight isolates using the MLST web server (www.genomicepidemiology.org).13 Two different MLST schemes were used: the Oxford MLST scheme and the Pasteur MLST scheme (http://pubmlst.org/abaumannii).

Identification of resistance genes

The ResFinder web server (www.genomicepidemiology.org) was used to identify acquired antimicrobial resistance genes in the assembled WGS data, using a threshold of 100% identity for the genes encoding β-lactamases and 98.00% identity for all other genes. ResFinder detects the presence of resistance genes, but not functional integrity and expression or resistance due to acquired variation in housekeeping genes.14

Results and discussion

Screening for blaOXA genes with multiplex PCR detected six isolates with blaOXA-23-like genes and two isolates with blaOXA-60-like genes (data not shown). ResFinder analysis of the WGS data identified blaOXA-23 in the six isolates with blaOXA-23-like genes and blaOXA-72 genes in the two blaOXA-40-like group isolates. Furthermore, blaOXA-66 (n=1), blaOXA-68 (n=5) and blaOXA-92 (n=2), all belonging to the intrinsic blaOXA-51 group, were identified (Table 1). To our knowledge, blaOXA-92 is rare and has only previously been reported from a clinical isolate from a patient in Greece.15

The eight isolates had four different PFGE profiles using Apal as restriction enzyme; four of the isolates had related types (B1, B2) (Table 1 and Figure S1, available as Supplementary data at JAC Online). PFGE using Smal as restriction enzyme revealed that these four isolates comprised two unrelated types (Figure S2).

Using the Pasteur MLST scheme, five of the eight isolates belonged to ST2 (CC2; where CC stands for clonal complex), two isolates to ST1 (CC1) and one isolate to ST158 (Table 1). The five isolates belonging to ST2 (using the Pasteur scheme) were further divided into ST195, ST208 and ST218 using the Oxford MLST scheme. Furthermore, ST499 and the new ST812 (double-locus variant of ST321) were identified for three isolates using the Oxford MLST scheme (Table 1). The Oxford MLST scheme had a higher discriminatory power than the Pasteur MLST scheme.
### Table 1. Description of eight *A. baumannii* isolates from Odense University Hospital

<table>
<thead>
<tr>
<th>Isolate no.</th>
<th>Sample date</th>
<th>Origin</th>
<th>Sample</th>
<th>PFGE ApaI</th>
<th>PFGE Smal</th>
<th>Pasteur MLST</th>
<th>Oxford MLST</th>
<th>SNP profile</th>
<th>Background</th>
<th>Patient’s link to other countries</th>
<th>Resistance profile&lt;sup&gt;a&lt;/sup&gt;</th>
<th>β-Lactamase</th>
<th>Genes encoding non-β-lactam resistance</th>
</tr>
</thead>
<tbody>
<tr>
<td>AMA 341</td>
<td>16-12-2012</td>
<td>D</td>
<td>E</td>
<td>158</td>
<td>499</td>
<td></td>
<td></td>
<td></td>
<td>travel to Egypt</td>
<td>1 bla&lt;sub&gt;OXA-23&lt;/sub&gt;, bla&lt;sub&gt;OXA-65&lt;/sub&gt;, bla&lt;sub&gt;GES-11&lt;/sub&gt;</td>
<td>aph(3&lt;sup&gt;′&lt;/sup&gt;)-VI-a, oac(6&lt;sup&gt;′&lt;/sup&gt;)-Ib-cr, aadB, mph(E), mrs(E), sul1, dfrA7</td>
<td>aadA1, aac(6&lt;sup&gt;′&lt;/sup&gt;)-Ib-cr, aph(3&lt;sup&gt;′&lt;/sup&gt;)-VI-a, aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, armA, strB, strA, mph(E), mrs(E), catB8, sul1, tet(B)</td>
<td></td>
</tr>
<tr>
<td>AMA 474</td>
<td>18-08-2013</td>
<td>C</td>
<td>F</td>
<td>2</td>
<td>218</td>
<td></td>
<td></td>
<td></td>
<td>hospitalized in Slovakia</td>
<td>1 bla&lt;sub&gt;OXA-23&lt;/sub&gt;, bla&lt;sub&gt;OXA-66&lt;/sub&gt;, bla&lt;sub&gt;ADC-25&lt;/sub&gt;</td>
<td>aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, armA, strA, strB, mrs(E), mph(E), tet(B)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AMA 441</td>
<td>09-06-2013</td>
<td>B2</td>
<td>G</td>
<td>2</td>
<td>208</td>
<td></td>
<td></td>
<td></td>
<td>hospitalized in Greece</td>
<td>2 bla&lt;sub&gt;OXA-23&lt;/sub&gt;, bla&lt;sub&gt;OXA-66&lt;/sub&gt;, bla&lt;sub&gt;ADC-25&lt;/sub&gt;, bla&lt;sub&gt;TEM-1D&lt;/sub&gt;</td>
<td>aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, armA, strA, strB, mrs(E), mph(E), tet(B)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AMA 495</td>
<td>14-09-2013</td>
<td>B2</td>
<td>G</td>
<td>2</td>
<td>208</td>
<td></td>
<td></td>
<td></td>
<td>same SNP profile as AMA 441</td>
<td>none</td>
<td>2 bla&lt;sub&gt;OXA-23&lt;/sub&gt;, bla&lt;sub&gt;OXA-66&lt;/sub&gt;, bla&lt;sub&gt;ADC-25&lt;/sub&gt;, bla&lt;sub&gt;TEM-1D&lt;/sub&gt;</td>
<td>aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, armA, strA, strB, mrs(E), mph(E), tet(B)</td>
<td></td>
</tr>
<tr>
<td>AMA 517</td>
<td>04-10-2013</td>
<td>A</td>
<td>NT</td>
<td>1</td>
<td>812</td>
<td></td>
<td></td>
<td></td>
<td>born in Serbia</td>
<td>1 bla&lt;sub&gt;OXA-72&lt;/sub&gt;, bla&lt;sub&gt;OXA-92&lt;/sub&gt;, bla&lt;sub&gt;TEM-1D&lt;/sub&gt;</td>
<td>aadA1, aac(6&lt;sup&gt;′&lt;/sup&gt;)-Ib-cr, aph(3&lt;sup&gt;′&lt;/sup&gt;)-VI-a, aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, catA1, sul1, tet(A)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AMA 525</td>
<td>14-10-2013</td>
<td>A</td>
<td>NT</td>
<td>1</td>
<td>812</td>
<td></td>
<td></td>
<td></td>
<td>same SNP profile as AMA 517</td>
<td>none</td>
<td>1 bla&lt;sub&gt;OXA-72&lt;/sub&gt;, bla&lt;sub&gt;OXA-92&lt;/sub&gt;, bla&lt;sub&gt;TEM-1D&lt;/sub&gt;</td>
<td>aadA1, aac(6&lt;sup&gt;′&lt;/sup&gt;)-Ib-cr, aph(3&lt;sup&gt;′&lt;/sup&gt;)-VI-a, aph(3&lt;sup&gt;′&lt;/sup&gt;)-Ic, catA1, sul1, tet(A)</td>
<td></td>
</tr>
<tr>
<td>AMA 520</td>
<td>11-10-2013</td>
<td>B1</td>
<td>I</td>
<td>2</td>
<td>195</td>
<td></td>
<td></td>
<td></td>
<td>born in Pakistan</td>
<td>1 bla&lt;sub&gt;OXA-23&lt;/sub&gt;, bla&lt;sub&gt;OXA-66&lt;/sub&gt;, bla&lt;sub&gt;ADC-25&lt;/sub&gt;</td>
<td>armA, strA, strB, mph(E), mrs(E), sul2, tet(B)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AMA 524</td>
<td>14-10-2013</td>
<td>B1</td>
<td>I</td>
<td>2</td>
<td>195</td>
<td></td>
<td></td>
<td></td>
<td>same SNP profile as AMA 520</td>
<td>none</td>
<td>1 bla&lt;sub&gt;OXA-23&lt;/sub&gt;, bla&lt;sub&gt;OXA-66&lt;/sub&gt;, bla&lt;sub&gt;ADC-25&lt;/sub&gt;</td>
<td>armA, strA, strB, mph(E), mrs(E), sul2, tet(B)</td>
<td></td>
</tr>
</tbody>
</table>

NT, non-typeable.

<sup>a</sup>Resistance profiles: profile 1, resistant to ampicillin/sulbactam, ticarcillin/clavulanic acid, piperacillin/tazobactam, ceftriaxone, cefotaxime, ceftazidime, cefoperazone, cefepime, aztreonam, imipenem, meropenem, lomefloxacin, ciprofloxacin, levofloxacin, trimethoprim/sulfamethoxazole, gentamicin, tobramycin, amikacin and tetracycline; and profile 2, as profile 1, except susceptible to trimethoprim/sulfamethoxazole.
The phylogenetic analyses showed that six of the eight Danish A. baumannii isolates clustered into three clusters each consisting of two isolates (Table 1). AMA 474 and AMA 341 did not cluster with other Danish or international isolates included in the study. The PFGE, MLST and SNP profiles discarded the suspicion of an outbreak consisting of all four isolates detected in October 2013, but strongly suggested a spread of carbapenem-resistant A. baumannii in three instances: between two patients (with AMA 517 and AMA 525) staying in the same room during hospitalization; between two patients hospitalized in the same ward (with AMA 520 and AMA 524); and between two patients living in the same nursing home (with AMA 441 and AMA 495) (Table 1). In addition to the blaOXA genes, 8–13 other resistance genes were identified from the assembled genomes. In silico analyses showed isolates sharing SNP profiles and Oxford MLST profiles had identical resistance gene profiles (Table 1). The isolates were resistant to almost all tested antimicrobial agents and patterns were consistent between the three sets of related isolates (Table 1).

MLST can be useful for investigation of global epidemiology. In the present study, three of the patients had been travelling abroad prior to OXA-carbapenemase detection and two persons were of foreign ethnicity living in Denmark. The ST158 (Pasteur MLST) isolate was detected in a patient who had been to Egypt prior to detection (Table 1). To our knowledge, ST158 is rare and has only been reported from Iowa, USA, but ST615 and ST618, which are single-locus variants of ST158, have been reported from Egypt (Pasteur MLST).

The patient with A. baumannii ST218 (Oxford MLST) had been to Slovakia prior to detection of the isolate; however, according to the Oxford MLST database, ST218 has only been reported from Japan. One of the two patients with ST208 (Oxford MLST) had been to Greece before detection, but, according to the Oxford database, ST208 has only been reported from Egypt, the USA and China.

One of the two patients with A. baumannii ST195 (Oxford MLST) was a Pakistani male living in Denmark. A. baumannii ST195 has previously been reported from several Asian countries. Furthermore, Karah et al. found a ST195 OXA-23-like producing A. baumannii in a patient from Thailand.

In conclusion, the SNP profile, Oxford MLST and the resistance gene profiles obtained from the WGS data were useful for typing of the eight carbapenem-resistant A. baumannii. WGS analysis, including SNP calling, MLST and resistance gene profiles, adds significant information for comparison and potential tracing of international-spreading clones and should replace PFGE typing.

Acknowledgements
Karin Sixhøj Pedersen at Statens Serum Institut and Elisa Knudsen at Odense University Hospital are thanked for their excellent technical assistance.

Funding
This work was supported by the Danish Ministry of Health and Prevention as part of The Integrated Surveillance of ESBL/AmpC Producing E. coli and Carbapenemase Producing Bacteria.

Transparency declarations
None to declare.

Supplementary data
Figures S1 and S2 are available as Supplementary data at JAC Online (http://jac.oxfordjournals.org/).

References