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ANTIHISTOMONAL EFFECT OF ARTEMISININ AND ARTEMISIA ANNUA EXTRACTS IN EXPERIMENTALLY INFECTED TURKIES.

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Summary

In the present investigation, previously established clonal cultures of Histomonas meleagridis and an infection model for histomonosis were used to evaluate the effect of five different Artemisia annua derived materials (i.e. dry leaves, artemisinin; and hexane, dichloromethane or methanol extracts). Successful in vitro inhibition of H. meleagridis clones were observed in all tested materials, except for the methanol extract. Whereas no activity against xenic bacteria accompanying the six H. meleagridis clones was observed. The dichloromethane extract of dry leaves (Ext-DCM) and artemisinin displayed the most promising antihistomonal properties and both were subsequently evaluated in a standardized experimental infection model in turkeys. Supplementation of Ext-DCM or artemisinin were commenced from day one of life and the birds were subsequently infected with clonal H. meleagridis at 2 weeks of age. Regardless of treatment, all infected turkeys showed severe clinical histomonosis and demonstrated severe typhlohepatitis.

Introduction

Infection with Histomonas meleagridis in poultry flocks has re-emerged since the ban of drugs providing effective prophylaxis and treatments (McDougald, 2005). Within recent years, efforts have been made to find alternatives to chemotherapeutics to combat histomonosis. Despite this, only a limited number
of *in vitro* studies on the effects of natural compounds on *H. meleagridis* are available (Zenner et al., 2003; Grabensteiner et al., 2007; Hauck & Hafez, 2007; Arshad et al., 2008; Grabensteiner et al., 2008; van der Heijden & Landman, 2008a). The situation is similar when it comes to evaluating the impact of natural compounds on histomonosis *in vivo* (Duffy et al., 2004; Duffy et al., 2005; Hafez & Hauck, 2006; Grabensteiner et al., 2008; van der Heijden & Landman, 2008b).

*Artemisia annua* has been used as an herbal remedy in traditional Chinese medicine for treatment of fevers, including malaria (Klayman, 1985). Artemisinin is one of the main active components of this medicinal plant and has been shown to be effective against various protozoa, including *Plasmodium* spp. (Qinghaosu Antimalaria Coordinating Research Group, 1979) and *Eimeria* spp. (Allen et al., 1997).

The aim of the present study was to examine the effect of a panel of extracts of *A. annua* leaves, as well as pure artemisinin, on *H. meleagridis in vitro and in vivo* using a standardized experimental infection model in turkeys (Hess et al., 2006a).

**Materials and methods**

*In vitro*

Six different clonal cultures of *H. meleagridis* (Hm2, Hm3, Hm4, Hm6, Hm7 and Hm18) were used to evaluate the effect of artemisinin and *A. annua* materials. Clonal cultures were established through micromanipulation (Hess et al., 2006b). Dry leaves from *A. annua* were finely ground before use. Artemisinin (purity > 99%) was obtained. Crude extracts from *A. annua* leaves were made using hexane, dichloromethane or methanol.

A test system previously described by Grabensteiner et al. (2007) was used. The concentration levels of the materials in the test cultures were: dry leaves, 5-40 mg/ml; artemisinin, 5-20 mg/ml; and hexane (Ext-HEX), dichloromethane (Ext-DCM) or methanol extracts (Ext-MeOH), 0.5-1.5 mg/ml. Protozoan growth and viability was evaluated after 24 and 48 hours of incubation at 40°C. The number of protozoa was ascertained using hemacytometer as described by Zenner et al. (2003).

In a similar setup, protozoa from *Histomonas meleagridis/Turkey/Austria/2922-C6/04* (Hm6) were used to determine the inhibitory properties of artemisinin (*IC*₅₀) (10⁻¹ - 4×10⁻⁴ µM). The *IC*₅₀ was enumerated by graphical extrapolation.

Bacteria present in the same monoeucaryotic *Histomonas* cultures were isolated using selective media. The antibacterial activity of the test materials were assessed using the disc diffusion method (Bauer et al., 1966; CLSI, 2008). Test solutions (20 µl) in concentrations identical to those in the first experiment were loaded onto empty Sensi-discs.
The data analysis and statistical calculations were made using one way ANOVA followed by Tukey’s multiple comparison test. \( P \)-values of \( \leq 0.05 \) were considered as significant.

**In vivo**

Based on the results from the *in vitro* experiments an animal experiment was set up in order to investigate the effect of artemisinin and Ext-DCM on a virulent clonal culture of *H. meleagridis* (*Histomonas meleagridis*/Turkey/Austria/2922-C6/04) (Hess *et al.*, 2006a) in turkeys.

Sixty-five day-old turkey poultcs were randomly split into five groups with equal gender ratio (Table 1). From the first day of life the birds had unlimited access to feed (details mentioned below) and water, except a feed restriction for 5 hours directly after the infection. All procedures performed on the birds were approved by the institutional ethics committee and licensed by the Austrian government (licence number 68.205/0103-II/3b/2011).

**Table 1: Overview of treatment groups**

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment</th>
<th>No. of birds</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Ext-DCM 0.2% in drinking water (day 1-15); 0.1% (day 16-)</td>
<td>15</td>
</tr>
<tr>
<td>II</td>
<td>Artemisinin 100 ppm in feed</td>
<td>15</td>
</tr>
<tr>
<td>III</td>
<td>Artemisinin 2600 ppm in feed</td>
<td>15</td>
</tr>
<tr>
<td>IV</td>
<td>Infection control None</td>
<td>15</td>
</tr>
<tr>
<td>V</td>
<td>Negative control None</td>
<td>5</td>
</tr>
</tbody>
</table>

Administration of artemisinin was done via feed and Ext-DCM via drinking water. Supplementation started at first day of life and was continued throughout the experimental period. The dosage was determined due to the obtained *in vitro* results and followed the protocol of Grabensteiner *et al.* (2008). Artemisinin (purity > 99%) was added to the diets in concentrations of 100 ppm and 2600 ppm. Ext-DCM from dried *A. annua* leaves was dissolved in DMSO and administered to group I as follows: from day 1-15 received 0.2% pure extract in drinking water; from day 16 onwards the concentration was decreased to 0.1% due to reduced water intake. Fourteen days old, the birds (group I-IV) were infected intraloacally with \( 3 \times 10^5 \) *Histomonas meleagridis*/Turkey/Austria/2922-C6/04 passage 22 (Hess *et al.*, 2006b). Birds of group III died or had to be euthanized before inoculation was undertaken.

Daily recordings of any adverse clinical signs, mortality, and feed and water consumption were obtained. Birds were weighed and had blood taken once a week. For monitoring the *H. meleagridis* excretion cloacal swabs were taken three times weekly according to the protocol described recently (Hess *et al.*, 2006a).
Turkeys that survived the challenge were killed at termination of the experiment at 5 weeks of age (Hess et al., 2006a). Euthanasia due to severe histomonosis or killing of chickens at specific time points was performed by intravenous anaesthesia with thiopental followed by bleeding.

Pathological examination was performed on all birds. Lesions indicative for histomonosis in the caeca and the livers were noted with scores ranging from 0 for no lesions to 4 describing the most severe lesions, according to recently described protocols (Windisch & Hess, 2010; Zahoor et al., 2011).

**Results**

*In vitro*

Addition of dry leaf plant powder resulted in complete growth inhibition at 20 and 40 mg/ml in Hm2, Hm3, Hm6, Hm7 and Hm18, whereas complete inhibition of cell proliferation was only seen at 40 mg/ml dry leaf powder in Hm4. The MLC for dry leaf powder was determined to 40 mg/ml after 24 h. For artemisinin, complete inhibition of protozoa proliferation was not observed in any of the six clones. However, significant reductions of histomonads were noticed after incubation for 48 h with artemisinin in all cultures. Growth of clonal cultures receiving 0.5 mg/ml Ext-HEX was significantly reduced in all six clones at 1.0 and 1.5 mg/ml Ext-HEX after 48 h. However, only 1.5 mg/ml resulted in complete inhibition in the six clonal cultures at 24 h of incubation. The MLC was determined to 1.5 mg/ml for Ext-HEX. When adding Ext-DCM to the protozoa cultures significant inhibitory effect against histomonads of all clones was observed at all three tested concentrations. Complete inhibition of the cultures after 24 h of incubation was observed at Ext-DCM levels of 1.0 mg/ml and 1.5 mg/ml (MLC= 1.0 mg/ml). In contrast to Ext-HEX and Ext-DCM, Ext-MeOH was not able to induce complete inhibition in any of the tested clonal cultures of *H. meleagris*.

The IC$_{50}$ for artemisinin after 48 h was determined to 4.586×10$^3$ µM, which equals 1.3 mg/ml in test solution.

No inhibitory effect of dry leaf powder, artemisinin, Ext-HEX, Ext-DCM or Ext-MeOH was observed in any of the 19 isolated bacterial strains from the six investigated *H. meleagris* clones.

*In vivo*

At day 5-7 following feeding, 7 out of 15 birds from group III died unexpectedly. The remaining birds of this group displayed increasing depression and anorexia and were therefore euthanized. Post mortem section findings were suggestive of intoxication.

Furthermore, it was observed that the water containing 0.2% Ext-DCM (group I) had a very pronounced herbal odour. At 2 weeks of age the water intake of those turkeys decreased to 60-70% compared to turkeys of the remaining
groups. For welfare reasons based on the lower water consumption the concentration of Ext-DCM was hereafter reduced to 0.1% in group I.

Except from one bird in group II, *H. meleagridis* was found in cloacal swabs from all birds (group I, II and IV) at least once during the experiment. No histomonads could be recovered from any of the turkeys in group V.

All infected turkeys (groups I, II, IV) showed various clinical signs of histomonosis, from general depression and ruffled feathers to sulphurous coloured diarrhoea and sudden death. Turkeys that died or had to be euthanized due to histomonosis displayed severe disease specific lesions in the caeca and livers. Furthermore, necropsy of the 3 surviving turkeys revealed severe lesions in caeca and livers similar to pathological changes of turkeys that died from the disease (Table 2).

**Table 2:** Mortality and median hepatic and caecal lesion scores (LS) of turkeys suffering from histomonosis.

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment</th>
<th>Mortality due to histomonosis</th>
<th>Lesions specific to histomonosis</th>
<th>Median LS</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>(No. of birds)</td>
<td>(No. of birds)</td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>0.2% (0.1%) Ext-DCM</td>
<td>13/15</td>
<td>13/13</td>
<td>4</td>
</tr>
<tr>
<td>II</td>
<td>100 ppm Artemisinin</td>
<td>13/15</td>
<td>15/15</td>
<td>4</td>
</tr>
<tr>
<td>IV</td>
<td>Infection control</td>
<td>13/14</td>
<td>14/14</td>
<td>4</td>
</tr>
<tr>
<td>V</td>
<td>Negative control</td>
<td>0/5</td>
<td>0/5</td>
<td>0</td>
</tr>
</tbody>
</table>

**Discussion**

The present *in vitro* experiment revealed significant dose dependant reductions in protozoal counts of all six tested clones of *H. meleagridis* for the tested concentrations of artemisinin and Ext-DCM. Although significant reduction rates were observed in all six clones, none of the artemisinin concentrations were able to induce a total inhibition of histomonad proliferation. Hence, no MLC could be determined. Based on the *in vitro* results, the IC$_{50}$ of artemisinin was determined to $4.586 \times 10^3$ µM, which is considerably higher than for malaria parasites, where IC$_{50}$ values are within the nanomolar range (Golenser *et al.*, 2006). Ext-DCM was the most effective leaf extract, displaying complete inhibition of protozoal multiplication at 1.0 mg/ml in all clonal *H. meleagridis* cultures.

At present histomonads need accompanying bacteria when cultured *in vitro*, however the role of the bacteria is not clear (McDougald, 2005). In the present study, the amount of artemisinin loaded onto the discs (100-300 µg/disc) had no antibacterial effect.

The results of the antiprotozoal screening and the antibacterial tests suggest that the observed inhibitory effect of dried *A. annua* leaves, artemisinin, Ext-HEX and Ext-DCM is due to a direct effect on histomonads. Ext-DCM and
artemisinin were found to have the strongest antihistomonal effect in the *in vitro* studies and were therefore selected for further *in vivo* testing.

Despite treatment with the Ext-DCM or artemisinin, the clinical outcome in turkeys was almost similar and of the same severity as noticed for the infected but untreated turkeys. Except for three birds, all infected turkeys died or had to be euthanized due to severe clinical signs.

An explanation for the discrepancy between *in vitro* and *in vivo* efficacy in the present investigation is unclear. It is possible that low bioavailability (Titulaer et al., 1990) may contribute to insufficient concentrations of artemisinins in the birds. However, no information on the bioavailability and metabolism is yet available for poultry or other avian species.

The post mortem findings from the administrated 2600 ppm artemisinin group (III) may be indicative of intoxication and further investigations are on-going.

**Conclusion**

Despite considerable *in vitro* antihistomonal properties, neither artemisinin nor Ext-DCM of *A. annua* were able to prevent experimental histomonosis in turkeys at the given concentrations. Thus, the results of this study clearly demonstrate the importance of defined *in vivo* experimentations in order to assess and verify *in vitro* results.

**Acknowledgements**

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**References**


CLSI (2008). *M31-A3 performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals; approved standard*


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