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Published in:
Frontiers in Plant Science

DOI:
10.3389/fpls.2012.00209

Publication date:
2012

Document Version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
Receptor-like kinase complexes in plant innate immunity

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Keywords: receptor-like kinases, complexes, plant immunity, signaling, defense

INTRODUCTION

Autotrophs, like plants, are the source of nutrients for heterotrophs. Plants are members of complex communities and have co-evolved commensal and pathological relationships with microbes. A fine balancing act is required to effectively combat invasion by pathogenic heterotrophs while effectively guarding resources for vegetative and reproductive growth (King and Roughgarden, 1982). This entails appropriately timed activation of defense responses to conserve energy for producing numerous healthy progeny, thus increasing evolutionary fitness through this adaptive plasticity (Sultan, 2000). Detecting harmful heterotrophs and converting this recognition to intracellular signals aimed at combating the invader and alerting surrounding tissue, is a major challenge, especially since pathogens co-evolve with their hosts to evade discovery (Frank, 1992; Lehti-Shiu et al., 2009).

Receptor-like kinases (RLKs) are surface localized, transmembrane receptors comprising a large family of well-studied kinases. RLKs signal through their transmembrane and juxtamembrane domains with the aid of various interacting partners and downstream components. The N-terminal extracellular domain defines ligand specificity, and RLK families are subclassified according to this domain. The most studied of these subfamilies include those with (1) leucine-rich repeat (LRR) domains, (2) LysM domains (LYM), and (3) the Catharanthus roseus RLK-like (CrRLK1L) domain. These proteins recognize distinct ligands of microbial origin or ligands derived from intracellular protein/carbohydrate signals. For example, the pattern-recognition receptor (PRR) AtFLS2 recognizes fgl2 from flagellin, and the PRR AtEFR recognizes elf18 from elongation factor (EF-Tu). Upon binding of their cognate ligands, the aforementioned RLKs activate generic immune responses termed pattern-triggered immunity (PTI). RLKs can form complexes with other family members and engage a variety of intracellular signaling components and regulatory pathways upon stimulation. This review focuses on recently emerging new data about how these receptors form protein complexes to exert their function.

THE LRR FAMILY

The plant RLK family has more than 600 members in Arabidopsis (Shiu et al., 2004). RLKs are divided into 44 sub-families depending on their N-terminal domains. While RLKs have been implicated in many biologically important processes (Gish and Clark, 2011), this review focuses on RLKs involved in pathogen detection.

RLKs involved in immunity are so-called pattern-recognition receptors (PRRs) that detect pathogen-associated molecular patterns (PAMPs) and, upon binding of their cognate elicitors, initiate a well-characterized set of defense responses termed PAMP-triggered immunity (PTI). Features of PTI include reactive oxygen species (ROS) production, callose deposition, generation of secondary messengers, and defense gene expression (Jones and Dangl, 2006). RLK elicitation also leads to activation of several mitogen-activated protein (MAP) kinases (Suarez-Rodriguez et al., 2007; Mithoe et al., 2011). PAMPs, and more broadly, microbial-associated molecular patterns (MAMPs) and damage-associated molecular patterns (DAMPs), can activate RLKs (Le rouge et al., 1990; Gomez-Gomez and Boller, 2000; Zapfel et al., 2006; Krol et al., 2010). Binding of PAMPs and DAMPs to their specific receptors leads to a broad range of downstream signaling events and effects. Figures 1A–C gives an overview of some of the complexes of Xa21, FS2, and EF-Tu receptor (EFR) that will be discussed in this review. Figure 1D shows biological effects of FLS2, Xa21 and EFR.

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et al., 2010a) and the RD kinase BAK1 (Chinchilla et al., 2007; Heese et al., 2007). These receptors present the core of our current knowledge regarding RLKs involved in defense.

**Xa21**

The Xa21 extracellular domain is composed of 23 LRRs and was one of the first eukaryotic RLKs found to be involved in resistance (Song et al., 1995; Wang et al., 2006). Xa21 binds the Xanthomonas oryzae pv. oryzae (Xoo) secreted tyrosine (Tyr) O-sulfonation peptide AxYS22 (Lee et al., 2009). Much has been learned about the function of Xa21. For example, the amino acids Ser686, Thr688, and Ser689 in the cytosolic JM domain are important for stability and for endoplasmic reticulum (ER) processing (Xu et al., 2006; Park et al., 2010a). Phosphorylation of residues in the JM domain is also critical for the activation of Xa21 and binding of at least four Xa21-binding proteins named XB3, XB15, XB24, and XB10 (OsWRKY62; Park et al., 2010b) associated with Xa21 via the JM domain. These interactions are all dependent on Thr705 since mutation of this JM domain residue abolishes XB-Xa21 binding (Chen et al., 2010a).

XB3 is an E3 ligase important for Xa21 accumulation and is a substrate for Xa21 kinase activity, although the biological relevance of this relationship is still unclear. After Xa21 binds AxYS22, XB3 is activated by transphosphorylation and likely leads to cleavage of a negative regulator of defense or even of itself, allowing other interactions to take place (Wang et al., 2006).

Xa21 is regulated by two proteins through phosphorylation; XB15, a protein phosphatase 2C (PP2C) and XB24, a protein with intrinsic ATPase activity (Park et al., 2008). XB15 dephosphorylates Xa21 and XB15 over-expression reduces Xoo resistance while xb15 null-mutants exhibit increased cell death and resistance to Xoo. This would point to a negative regulatory role of XB15. On the other hand, XB24 promotes autophosphorylation of Xa21 and may be required to prevent proteolytic cleavage of Xa21. The complex between XB24 and Xa21 dissociates upon Xoo infection or AxYS22 binding (Chen et al., 2010b). Phosphorylation, especially in the JM domain, plays a critical role in Xa21 stability. It is clear that autophosphorylation of certain residues in Xa21 promotes an inactive state but the exact changes in phosphorylation status upon pathogen infection remain largely unknown.

Xa21 binds to the WRKY transcription factor XB10 and this binding requires an active Xa21 kinase domain. Binding of the AxYS22 peptide to Xa21 leads to translocation of a Xa21 kinase domain-GFP fragment to the nucleus where it interacts with XB10.
AxYS22 ligand is perceived, the majority of the receptor is found in the ER. August 2012 | Volume 3 | Article 209 | www.frontiersin.org

Flagellin-derived peptide flg22 has been shown using 125I-labeled flagellin to interact with BiP3, an HSP70-like ATPase located in the ER, and to interact with other known BAK1 interactors such as BRI1, BKK1, SERK1, and SERK2. FLS2 interactions with SERK1, SERK2, and BKK have been detected, but its predominant association is with BAK1. BAK1 and BKK1 are thought to act cooperatively in PAMP signaling and resistance to biotrophic pathogens (Veronese et al., 2011). BAK1 and FLS2 also interact with Botrytis-induced kinase 1 (BIK1), which is a receptor-like cytoplasmic kinase (RLCK) implicated in resistance to necrotrophic pathogens (Veronese et al., 2006). BAK1 and FLS2 phosphorylate BIK1 (Lu et al., 2010) and BIK1 in turn phosphorylates both FLS2 and BAK1. This is thought to be an important signal amplification mechanism. However, since FLS2 has been shown to have very low catalytic activity in vitro (Schwessinger et al., 2011), BAK1 probably possesses the predominant kinase activity influencing BIK1 phosphorylation. The BIK1–FLS2/BAK1 association is decreased after flg22 sensing, suggesting that BIK1 is released to activate downstream signaling components (Lu et al., 2010). BIK1's role in PTI is dependent on complex interactions with major immune response regulators and may thus provide RLK signaling complexes with the ability to discriminate between biotrophic and necrotrophic pathogens (Laluk et al., 2011). Importantly, bir1 mutants display enhanced susceptibility to Pto DC3000, reduced flg22 responsiveness, as well as compromised flg22-induced resistance to virulent Pto DC3000. The BIK1-related kinases, PBS-like kinase 1 (PBL1) and PBL2 also interact with FLS2 and BAK1. pbl1 mutants show less reduction in PTI responses but the effect seems to be additive to BIK1 function (Zhang et al., 2010).

BAK1, BKK1, SERK1, and SERK2 have also been shown to interact with BIR1 (BAK1-interacting receptor-like kinase 1), an active protein kinase. The bir1 mutant exhibits increased resistance to biotrophic Pto DC3000 and HyaIoperonospora arabidopsidis Noco2, due to apparent R protein activation (Wang et al., 2011). The bir1 phenotype is partially rescued in bir1 pad4 double mutants, and is completely rescued in the bir1 pad4 sob1 (suppressor of bir1-1) triple mutant. Phytoalexin deficient 4 (PAD4) is one of the critical components required for Toll/interleukin-1 receptor (TIR) R protein signaling. Many constitutively active defense phenotypes that result from activated TIR R proteins are suppressed by PAD4 loss of function (Wiermer et al., 2005; Palma et al., 2010; Zhang et al., 2012). The aforementioned results thus indicate that bir1 is not responsible for BR-mediated resistance induction in bir1 occurs through SOBIR1. SOBIR1 is also a R-like receptor, and over-expression of SOBIR1 leads to activation of cell death. SOBIR1 does not function in flg22 sensing and does not interact with BIR1. Exactly how loss of BIR1 activates SOBIR1 is a mystery (Gao et al., 2009), and it is still uncertain whether BIR1 has a role in the PAMP signaling pathway.

Kinase-associated protein phosphatase (KAPP) interacts with the FLS2 kinase domain (Gomez-Gomez and Boller, 2000), and this interaction may be important for receptor endocytosis upon activation as was found for Aserk1 (Shah et al., 2002). KAPP has also been found in complexes with other RLKs (Williams et al., 1997; Stone et al., 1998) but whether it functions as a general regulator of a broader spectrum of RLKs needs to be explored.

FLS2 also interacts with E3 ligases that polyubiquitinate the receptor after flg22 signaling. FLS2 is subsequently degraded by the proteasome, which might constitute a mechanism for attenuation as has been described for the mammalian Toll-like receptor 4 (TLR4) and TLR9 (Chuang and Ulevitch, 2004). Plant U-Box 12 (PUB12) and PUB13, both E3 ubiquitin ligases, have been shown to be BAK1 phosphorylation targets, and this modification is required for their association with FLS2. This phosphorylation is reminiscent of the previously mentioned Xa21 phosphorylation of XB3. PUB12 and PUB13 control flg22-dependent, proteasome-mediated degradation of FLS2.
nucleotide-activated Ca2+, but similarly to Xa21, N-glycosylation is critical for EFR (et al., 2010). Given the many parallels between FLS2 and EFR, it has been found for EFR (Nekrasov et al., 2009; Saijo et al., 2009; Häweker et al., 2010). However, FLS2 has recently been shown to interact with the ritulin-like proteins RTNLB1 and RTNLB2. RTNLB1/2 are altogether involved in regulating FLS2 transport from the ER to the plasma membrane (Lee et al., 2011). In addition, stomatal cytokinesis defective 1 (SCD1) was identified by mass spectrometry as an FLS2 interaction partner. scd1 mutants display SA-dependent enhanced resistance to infection with Pto DC3000, as well as enhanced accumulation of PR1 transcripts and hydrogen peroxide. However, the same mutants are less sensitive to PAMPs, with reduced seedling growth inhibition and ROS production in response to flg22 or elf18 (Koraisik et al., 2010).

**EF-Tu RECEPTOR**

EF-Tu receptor is a LRR-RLK that recognizes the peptide elf18 from bacterial elongation factor (EF)-Tu. EFR and BAK1 have also been shown to interact in a ligand-dependent manner (Roux et al., 2011). Indeed, many of the signaling components downstream of EFR and FLS2 are shared. While both EFR and FLS2 are capable of associating with all members of the SERK family, BKK1, SERK1, SERK2 have a stronger association with EFR than with FLS2 (Roux et al., 2011). This might allow EFR to avoid pathogen effector action on the single SERKs. Studies of SERK function have been difficult due to their apparent redundancy and the lethality of some double mutants such as serk1, serk2 and bak1-1/4 bakk-1-1 (Colombolet et al., 2005; He et al., 2007). However, the discovery of a novel allele of bak1, bak1-5, enabled study of non-lethal bak1-5 bak1-4 double mutants. This revealed that BAK1 and BKK1 act cooperatively in PAMP signaling (Roux et al., 2011; Schwessinger et al., 2011). BAK1 is phosphorylated upon elf18 and flg22 treatment (Lu et al., 2010). Given the many parallels between FLS2 and EFR, it is possible that transphosphorylation of the EFR/BAK1 complex also occurs, although direct proof is still lacking. In contrast to FLS2, but similarly to Xa21, N-glycosylation is critical for EFR function and EFR is subject to ER quality control that requires several chaperones involved in ERQC for full activity (Häweker et al., 2010).

**PEPR1**

In contrast to the three receptors described above, Pep1 receptor 1 (PEPR1) binds AtPep1 (Yamaguchi et al., 2006) a DAMP derived from the precursor gene. PROPEP1, PEPR1 and PEPR2 act redundantly to perceive AtPep1. BAK1 was shown to interact with PEPR1 like FLS2 and EFR (Postel et al., 2010). PEPR1 possesses a putative guanylyl cyclase (GC) domain and cGMP production by the purified RLK was shown in vitro (Q. et al., 2010). Interestingly, a GC domain is also present in BR11 and was shown to have a catalytic function in vitro (Kozeri et al., 2007). This cGMP generated after elicitation may trigger a cyclic nucleotide-activated Ca2+ channel as part of its signaling activity (Ali et al., 2007).

**LyM FAMILY**

Chitin elicitor receptor kinase 1 (CERK1) is the best studied Arabidopsis LyM-RLK (Kaku et al., 2006; Miya et al., 2007; Wan et al., 2008), and direct binding of chitin to CERK1 has been detected (Iizasa et al., 2010; Petutschnig et al., 2010). Unlike FLS2 and EFR, CERK1’s perception of fungal chitin is BAK1-independent. In rice, Chitin elicitor-binding protein (CdbBP), a LyM domain-containing protein, associates with OsCerk1 and these proteins function together in a hetero-oligomer receptor complex to elicit chitin signaling in a ligand-dependent manner (Shimizu et al., 2010). Two LyM domain proteins, LYM1 and LYM3, have recently been shown to be important for peptidoglycan (PGN), but not chitin recognition. LYM1 and LYM3 are not functionally redundant, and it has been proposed that LYM1, LYM3 and CERK1 may form a complex or complexes. cerk1 is hypersensitive to Pto DC3000 and shows reduced sensitivity to PGN, phenocopying lym1/lym3. However, CERK1 does not bind to PGN. Further, given the fact that neither LYM1 nor LYM3 contain a cytokinotic domain, a LYM1/LYM3/CERK1 complex seems likely (Wilmann et al., 2011). RLKs often hetero-oligomerize for optimal functioning as seen in the co-operativity of FLS2/BAK1, EFR/BAK1 and PEPR1/PEPR2.

**CRLK1 FAMILY**

Another RLK, FERONIA (FER) was first shown to control pollen tube reception (Escobar-Restrepo et al., 2007). However, the expression of FER throughout the plant suggests a general function not strictly associated with root development or pollen tube reception. Indeed, FER has more recently been shown to aid powdery mildew (PM) penetration into host cells (Kessler et al., 2010) and to be responsible for susceptibility to the oomycete H. arabidopsidis (Niba and Cheung, 2011). It is suspected that FER might play a role in controlling localization of MLO family proteins, known to be important for PM infection (Cionini et al., 2006), as it does for NTA during pollen tube reception. This however still needs to be shown, as well as whether ROS signaling has an effect on MLO localization. Given the many roles of FER it is not surprising to find that it is important for disease resistance as well. FER appears to exert is signaling functions by controlling ROS production. FER was shown to interact with guanine nucleotide exchange factors (GEFs) that regulate RHO GTPases (RAC/ROPs). RAC/ROP is known to play important roles in stress-induced responses. In rice, the binding of a RAC/ROP called Rac GTase to NADPH oxidases has been characterized, and Rac GTase was shown to be required for ROS production (Wong et al., 2007). In Arabidopsis, Rop2 was shown to aid immunoprecipitate with FER. In addition, transgenic plants expressing constitutively active, GTP-bound Rop2 displayed increased ROS production (Cheung and Wu, 2011). This indicates that a FER-GEF-RAC/ROP complex is likely able to affect ROS production. While ROS play a role in root development, there are hints that FER is involved in ROS production during PAMP signaling in leaves. For example, FER is enriched in detergent-resistant membranes (DRMs) after flg22 treatment, and FER shows flg22-induced phosphorylation (Benschop et al., 2007). FER mutants also exhibit enhanced ROS production, and aberrant stomatal responses upon flg22 treatment (Keinath et al., 2010). The increase
in ROS production in the fer mutant is puzzling given the reduced Rho GTPase activity in this mutant (Duan et al., 2010). The relationship between FLS2 and FER in the control of ROS production is very interesting and should attract attention in the near future.

CONCLUDING REMARKS

There have been enormous advancements in our knowledge about RLRK signaling in the last decade, but many questions still remain unanswered. For example, the link between the PRR receptors and innate immunity. Plant Cell 19, 1081–1095.

REFERENCES

Albrechtsen, C., Breuton, F., Sevag,
zuc, C., Schmeisser, B., Gmeiner-
inhibit pathogen-associated molec-
ular-pattern-triggered immunity indepen-

Ali, R., Ma, W., Lemtiri-Chlieh, F.,
\n
CHINCHILLA, D., Bauer, Z., Regenass,


Coombe, C., Humphry, E. M., Hurst,

dropes for function of the immune

www.frontiern.org

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Frank, S. A. (1992). Models of plant-


Gou, X., Yuan, T., Bi, D., Cheng,
Y. T., Chou, S., Li, X., and Zhang, Y. (2010). Regulation of cell death and innate immunity by two receptor-like


Jones, J. D. G., Felix, G., and Boller,

Rho GTPase activity in this mutant (Duan et al., 2010). The

Greeff et al. Receptor-like kinases


Benschop, J. J., Mohamed, S., O’Hafferty, M., Hock, A. J. R., Si-

not, M., and Monka, F. I. H. (2007). Quantitative phosphoproteomics of 

casual elicitor signaling in Arabidopsis.

Macell Proteomics 6, 1190–1214.

Chen, X., Chen, M., Calkin, P. E., Jiang, C., Evans, D., Cao, F., and Ronald, P. C. (2010a). A conserved threonine residue in the jumonji domain of the XA21 pat-


Chen, X., Chen, M., Calkin, P. E., and Ronald, P. C. (2010b). An AtGTPase pro-


Chou, A. Y., and Wu, H.-M. (2010). FER-

onial pattern)-induced changes in

pan-protein ligase regulating Toll-like


Kong, D., and Roughgarden, J. (1982). Multiple switches between repre-


cytide 1 involves the pattern recogni-

sis xylanase brassinosteroid receptor (ABRE1) contains a domain that functions as a guanylyl cyclase in vitro. Plant Cell 2, 4485–4500.

ments for function of the immune


Jones, J. D. G., Felix, G., and Boller,

and production of ROS and activation of MAP kinase is still coming. Nevertheless, a quite comprehensive picture of the route from receptor activation to enhanced defense gene expression has emerged for XA21 and similar data for FLS2 and FER are sure to come to light.

ACKNOWLEDGMENTS

This work was supported by grants to Morten Petersen from the Danish Research Council for Technology and Production (11-106302) and the Strategic Research Council (09-067148).


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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 03 May 2012; paper pending published: 03 June 2012; accepted: 16 August 2012; published online: 24 August 2012.


This article was submitted to Frontiers in Plant Proteomics, a specialty of Frontiers in Plant Science.

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