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Long-Chain Polyunsaturated Fatty Acids in Breast-Milk and Erythrocytes and Neurodevelopmental Outcomes in Danish Late-Preterm Infants

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Abstract

Background: The supply of long-chain polyunsaturated fatty acids (LC-PUFA) during pregnancy and early lactation has been shown to affect cognitive development in preterm infants, but the effect on early neurodevelopment of late-preterm infants has not yet been examined.

Aim: To examine the fatty acid composition of late-preterm human milk and identify possible associations between infant LC-PUFA status and perinatal as well as 1-year neurobehavioral outcomes.

Methods: Mother’s milk and erythrocytes (RBC) were sampled from 53 Danish late-preterm infants (33-36 weeks of gestation) 1 week and 1 month after delivery, and 3 months corrected age. Fatty acid composition was determined by gas-liquid chromatography. Neurodevelopmental outcomes were assessed by the Nicu Network Neurobehavioral Scale (NNNS) at 1 week and 1 month and the Bayley Scales (BSID-III) at 1 year corrected age.

Results: We found that breast-milk content of arachidonic acid (AA) and docosahexaenoic acid (DHA) was similar to reported fatty acid compositions of term human milk. Infant RBC-AA decreased from 1 week to 1 month of age and the size of the decrease was associated with better NNNS-scores at 1 month, specifically on regulation (p=0.03). Infant RBC-AA at 1 month was also associated with a lower 1-year corrected age BSID-III score of receptive language (p=0.05) and fine motor development (p=0.03). Infant RBC-DHA did not decrease significantly after delivery and was not associated with any of the developmental outcomes.

Conclusion: Breast-milk LC-PUFA content was reflected in the RBC LC-PUFA status of the infant. Early RBC-AA status was associated with both early and long-term neurobehavioral development, but not in a consistent way.

Keywords: Late-preterm infant; Long-chain polyunsaturated fatty acid; Neurodevelopment; Breastfeeding; Human milk; Erythrocytes

Abbreviations: AA: Arachidonic Acid; BSID-III: Bayley Scale of Infant Development, Third Edition; DHA: Docosahexaenoic Acid; LA: Linoleic Acid; LC-PUFA: Long-chain Polyunsaturated Fatty Acid; NNNS: Nicu Network Neurobehavioral Scale; PUFA: Polyunsaturated Fatty Acid; QoM: Quality of Movement; RBC: Erythrocyte (Red Blood Cell); SD: Standard Deviation

Introduction

Long-chain polyunsaturated fatty acids (LC-PUFA), especially docosahexaenoic acid (C22:6n-3, DHA) and arachidonic acid (C20:4n-6, AA), may affect the development of the infant brain as well as the infant growth pattern both before and after delivery [1,2]. These LC-PUFAs accumulate in the brain alongside foetal body fat, particularly during the third trimester of pregnancy and the first few months post-term. They are found especially in the synaptic membranes of neurons of the cortical and midbrain grey matter [1,3] in areas responsible for sensory integration, motor control and attention [4]. During the third trimester the foetus receives approximately 50 mg/(kg*day) of total n-3 LC-PUFA and 400 mg/(kg*day) of total n-6 PUFA [5-7]. AA seems to accumulate to a larger extend than DHA in the foetal brain until around the time of expected delivery; but subsequently DHA is the dominant LC-PUFA [5,8].

Metabolic maturation of cells and organs is believed to take place in the late stage of gestation [9,10] and in the early months after delivery at which time the foetal brain undergoes several structural changes. Neural differentiation, formation and adaptation of synaptic functions as well as glial cell production and myelination take place alongside neuronal regression and conversion of the foetal cortical subplate into a mature cortical structure. This transition is believed to mediate the simultaneous change in infant neuro-behaviour, e.g. in motor control from primitive reflexes and jerky movements to smooth and volitional motion [5,11]. Preterm delivery carries a risk of disrupting these processes, and apparently healthy, late-preterm infants (defined as infants born with a gestational age of 34-36 weeks) can experience long-term developmental problems such as difficulties with attention span, language skills and visuo-spatial motor function [11].

Human milk displays large individual and regional variations in the content of LC-PUFA, most pronouncedly in DHA, which also varies during the course of the day reflecting the content of the individual meals of the mother [12]. Individual factors such as maternal lifestyle, dietary intake and supplementation are determinants for the presence of individual PUFA in the milk [1]. LC-PUFA has been shown to decrease during the lactation period, and breast-milk of mothers with preterm delivery has been shown to have a higher content of LC-PUFA.
been shown to correlate with outcome measures on the Bayley Scales of Infant Development (BSID) [21,22]. NNNS is designed for use within gestational age 30 to approximately 46–48 weeks and normative preterm and healthy full term samples are available in the test manual [23]. For this study, we selected 3 out of 13 available summary scores as described in the Statistics section below.

Infant developmental milestones were recorded in the period between 3 and 12 months corrected age. Developmental outcomes at 12 months corrected age were determined by the third version of the BSID (III, Danish version 2009, Pearson Assessment, San Antonio, USA). BSID is the long-standing gold standard for infant global developmental testing and a proven method for detecting developmental anomalies. BSID evaluates cognitive, language and motor skills. Receptive and expressive language skills are evaluated separately and combined into an aggregate score and the same applies to fine motor and gross motor skills [24].

Testing conditions for both NNNS and BSID were selected to create the best possibilities for the infants to stay relaxed and attentive. All examinations took place in a quiet ambulatory room without wall decorations. The curtains were closed and the computer screen turned off, and for the NNNS test electric lights were off to keep the infants from focusing on lights and shadows. Both tests were performed halfway between meals if possible and the infants were examined in the presence of only one parent to avoid distraction. Furthermore, we aimed to begin the NNNS examination while the infant was still asleep. All examinations were video recorded, and performed by the lead investigator after proper training by a certified instructor [10].

Blood sampling and fatty acid analysis

At each visit, blood was drawn in 2-mL lithium-heparin vials. Erythrocytes (RBC) were immediately isolated by centrifugation at 2300 × g for 5 minutes at 4°C. Plasma was removed and the RBC were washed three times in 0.9% NaCl with 1 mM EDTA at pH 7.4, re-suspended 1:1 in an isotonic NaCl-solution with a final concentration of 0.1% butylated hydroxytoluene (VWR – Bie & Berntsen, Herlev, Denmark) and stored under N2 at -80°C for a maximum of 3 months.

RBC fatty acid composition was determined as previously described [25]. In brief: RBC were haemolysed in distilled water, and the lipids were extracted by the Folch procedure [26]. Fatty acid methyl esters were produced from the extracted RBC lipids using a BF3-catalyzed method [27] and extracted with heptane and analysed by gas-liquid chromatography on an Agilent 6890 Series II chromatograph (Agilent Inc., Santa Clara, California, USA) equipped with a flame ionization detector and a SP2380 capillary column (length 60 m; internal diameter 0.25 mm and film thickness 0.2 µm from Supelco Inc, Pennsylvania, USA) [28]. Fatty acid peaks were produced from the extracted RBC lipids using a BF3-catalyzed method [27] and extracted with heptane and analysed by gas-liquid chromatography on an Agilent 6890 Series II chromatograph (Agilent Inc., Santa Clara, California, USA) equipped with a flame ionization detector and a SP2380 capillary column (length 60 m; internal diameter 0.25 mm and film thickness 0.2 µm from Supelco Inc, Pennsylvania, USA) [28]. Fatty acid peaks were produced from the extracted RBC lipids using a BF3-catalyzed method [27] and extracted with heptane and analysed by gas-liquid chromatography on an Agilent 6890 Series II chromatograph (Agilent Inc., Santa Clara, California, USA) equipped with a flame ionization detector and a SP2380 capillary column (length 60 m; internal diameter 0.25 mm and film thickness 0.2 µm from Supelco Inc, Pennsylvania, USA) [28].

Milk sampling and fatty acid analysis

A 50 mL breast-milk sample was collected between 5 and 8 a.m. in the last days up to each examination using an electric hand-held breast pump (Medela Swing, Medela AG, Baar, Switzerland). Many mothers could not deliver the 50 mL at once, especially at the first sampling, so they were allowed up to three consecutive mornings to collect
the sample. Mothers were instructed to freeze their milk samples immediately after pumping in their home freezer, and in case they had to do the pumping on more than one morning, to keep the samples in separate containers. The mothers transported the milk samples from their home freezer to the clinic in an UV-proof black cooler bag containing a freezing element with capacity to keep the bag content frozen for the duration of the drive to the clinic. Immediately upon arrival, milk samples were placed at -18°C until further processing. Within a few days (≤7 days), the samples were thawed in a 20°C water bath; sub-samples were pooled and gently mixed. The samples were aliquoted in 0.5 mL containers and 0.1% butylated hydroxytoluene in ethanol was added to aliquots for fatty acid analysis. The samples were then blown with N2 and kept at -80°C until analysis. Samples were analysed within 3 months from collection.

The breast-milk samples were freeze-dried for analysis of fatty acid composition. The freeze-dried material was mixed with 1 mL of heptane containing 0.4 mg/mL of internal standard (C12:1 cis11 triacylglycerol from NuChekPrep Inc, Elysian MN, USA). Methylation was carried out as a two-step procedure based on Jenkins [29] with modifications. Base-catalysed methylation carried out by addition of 0.2 mL 25% sodium methoxide, followed by incubation at 50°C for 10 min. After cooling 1.5 mL of a 10% methanolic HCl was added and the samples were incubated at 90°C for 30 min. After cooling 3 mL 10% K2C03 was added, tubes were centrifuged and the heptane layer was isolated. Separation of the fatty acid methyl esters was carried out on an Agilent 6890 gas chromatograph with a flame ionization detector and a Restek 2560 capillary column (length 100 m, internal diameter 0.25 mm and film thickness 0.20 µm from Restek, Bellefonte, Pennsylvania, USA). The temperature program started at 100°C for 5 min, followed by an increase of 3°C/min to 140°C, another increase of 20°C/min to 160°C, where it was kept for 20 min followed by an increase of 12°C/min to 220°C, which was then kept for 15 min after which it was increased at 8°C/min to a final temperature of 240°C for 8 min. The carrier gas was helium at a constant flow of 1 mL/min. The individual peaks were identified by use of external standards (37 component mix from Supelco, Belafonte, USA and GLC 469 standard from NuChekPrep Inc., Elysian, MN, USA) and quantified by use of the internal standard. The total sum of fatty acids was calculated as g/L milk and the content of individual fatty acids was expressed as mass-% fatty acids. The coefficients of variation for LA, AA and DHA were 4%, 5% and 5%, respectively.

Statistics

The statistical analyses were performed by IBM SPSS Statistics software (version 20.0). Data are presented as frequencies or mean ± SD. Students t-test was used for group comparisons e.g. gender differences and participants vs. non-participants (excluded infants, those refusing to participate and those lost to follow-up). Correlations between exposures, outcomes and possible covariates were calculated using the non-parametric Spearman’s test. Assumption of normality was tested by Shapiro-Wilk’s test and visual inspection of histograms, and by examining the residual plots of the final univariate linear models.

Univariate general linear models were used to test associations between LC-PUFA and developmental scores. The models were built by backwards-stepwise exclusion (p<0.05) of possible covariates: gender, gestational age at delivery, maternal education, socioeconomic status (composite score of education and occupation as defined by The Danish National Centre for Social Research) maternal psychiatric disorder (e.g. depression), multiple pregnancy, smoking in pregnancy, maternal multiparity and age at neurological assessment (days). None of these covariates were correlated (Spearman p>0.05). Furthermore, maternal smoking was also tested as a potential covariate for breast-milk LC-PUFA levels.

To avoid multiple testing we carefully selected the measures of breast-milk and infant RBC LC-PUFA as well as the NNNS scores to be used in analyses of association. In order to do that we used information about breastfeeding from questionnaires and interviews and data on the fatty acid composition of the breast-milk and infant RBC samples to determine which samples to use in our correlation and association analyses. The individual variation in breast-milk DHA (1.03% at 1 week and 1.35% at 1 month) exceeded the changes over time, i.e. between the means at 1 week and 1 month (0.1%), so we estimated that the mean DHA of all three samples would give the most accurate description of infant DHA intake during the breastfeeding period. We tested this assumption by comparing the mean after 1 month with the combined mean (by t-test), and as we found no difference, we chose the combined mean as our independent variable in the subsequent analyses. The same rationale applied to breast-milk AA.

Infant RBC LC-PUFA levels are expected to be associated with brain accumulation of AA and DHA [30] so we considered infant RBC levels the best proxy for brain LC-PUFAs and chose RBC as our main focus in the analysis of association with infant neurodevelopment. The following rationale was applied in selecting which RBC samples to use in our analyses: Because of the approximately 3-months turnover of the RBC, the 1-week infant RBC sample reflects the LC-PUFA supply during pregnancy and does not reflect supply from breastfeeding. The 1-month sample reflects the LC-PUFA supply from 32 weeks of gestation to 4 weeks post-term, and thus covers 8-12 weeks of extrauterine enteral feeding depending on the infant’s gestational age at delivery, whereas the sample taken at 3 months corrected age in theory should reflect the entire early feeding period. However, as descriptive statistics showed an increasing uniformity of the 3 months samples and a discrepancy between the 3 months sample and the previous two samples and as an increasing number of children stopped breastfeeding, we considered the sample taken at 1 month post-term to be the best measure of infant LC-PUFA uptake from breast-milk.

NNNS generates 13 summary scores, some of which measure primarily physiological instability or neurological damage, which was outside the scope of this study (excitability, lethargy, stress/abstinence, handling, hyper-/hypotonia [31]). To find the scores most suitable to use for comparison of development between infants, we reviewed the internal correlation of scores over time (by Spearman’s test), number of missing values, and normal distribution as well as how the scores reflected hypotheses in previous studies in the LC-PUFA field. Three out of the remaining 7 summary scores were used in our analyses: quality of movement (QoM), regulation and arousal (which was strongly negatively correlated with habituation, r=−0.42, p=0.006). QoM summarizes motor maturity, control and activity as well as the ability to perform smooth movements without startles, tremors or jitters. Regulation evaluates the combined scores of attention, motor and psychological activity and the infant’s capacity to regain comfort with and without examiner assistance. Arousal is a measure of the infant’s overall level of motor activity and irritability, fuss and cry in response to handling, as well as the state of arousal reached during the examination [32]. Higher scores in all three variables indicate a higher degree of neuro-maturation.

We examined if RBC LC-PUFA status during the perinatal period was associated with the three NNNS measures at delivery and at 1
month post-delivery by correlation analysis and univariate linear regression models adjusted for gender and gestational age. BSID-III scores at 1 year corrected age (cognitive, receptive language, expressive language, fine motor and gross motor scale scores) were also tested against AA and DHA in breast-milk and infant RBC. To determine whether neuro-developmental status at delivery excerts a confounding influence on developmental status in later infancy, we included the NNNS measures as covariates in the final models for the BSID-III outcomes.

Due to the explorative nature and the small sample size of this study, performing a rigorous correction for multiple testing (e.g. by Bonferroni) would increase the risk of type-II errors considerably. We therefore chose not to correct for multiple testing, but to report the p-values of our analyses and to discuss the potential relevance of findings with p-values <0.05.

Results

The inclusion rate was 69% (Figure 1). The characteristics of the included mothers and infants (Table 1) differed only from non-participants by slightly younger mothers (29±5 years vs. 31±4 years, p=0.048). The rate of loss to follow-up was 9% (n=5) and the lost infants differed from the remaining infants by a higher rate of maternal psychiatric disorder (p=0.015), although this was caused by only one twin mother (n=2).

Sixteen infants (30%) were never exclusively breastfed and introduction of complementary feeding was initiated at a mean age of 147±25 days corresponding to 116±24 days post-term. Infants who were breastfed at 3 months corrected age differed from those who were not by the frequency of maternal smoking in pregnancy (50% smokers in the non-breastfeeding group vs. 5.4% in the breastfeeding group, p<0.001) and maternal smoking was correlated with low socioeconomic status (r=0.29, p=0.035).

Fatty acid composition of breast-milk and infant RBC

The LC-PUFA composition of the breast-milk and the infant RBC samples are shown in Table 2. There was a significant 26% decline (p<0.001) in breast-milk DHA during the first month, but there was no significant difference between the fatty acid composition of the milk samples at 1 month after delivery and at 3 months corrected age. The mean levels of AA and DHA in the combined samples were 0.61±0.13 mass-% for AA and 0.66±0.28 mass-% for DHA. The RBC content of LA and DHA in the infants increased over time, whereas that of AA decreased (all changes p<0.05).

Overall, the correlations between breast-milk and infant RBC were positive for both AA and DHA (Table 3). Breast-milk and RBC-LA were only correlated at 1 month after delivery. There was no association between gestational age at delivery and infant RBC-DHA, AA or LA, although AA showed a trend towards a negative association (B=-0.07±0.032, p=0.051). There was a decrease in the variance for infant RBC-DHA and AA at 3 months corrected age, and the correlations between the infant RBC samples at 1 month and 3 months as well as between breast-milk and infant RBC at 3 months were not significant except for AA (Table 3), which may be due to the increase in formula feeding frequency after 1 month of age.

LC-PUFA and NNNS

Paired t-tests between the NNNS scores at 1 week and 1 month revealed that regulation and arousal increased over time, but no increase was seen in QoM (Table 5A). QoM at 1 week and 1 month of age were found to correlate (Spearman r=-0.31, p=0.02), whereas no correlations were seen for regulation (r=0.02, p=0.90) and arousal (r=0.12, p=0.39). There was no gender difference in the NNNS scores at any of the examinations.

There were no significant correlations (Spearman) between the levels of AA or DHA in infant RBC at 1 month of age and any of the NNNS scores. There was also no correlation between the changes in infant RBC-AA and DHA between 1 week and 1 month and the corresponding changes in the NNNS scores, but arousal at 1 month of age tended to correlate negatively with the change in infant RBC-AA (r=-0.30, p=0.050). The relationships between infant RBC LC-PUFA status and the NNNS neuro-developmental measures at 1 week and 1 month were also examined by analyses adjusted for gender and gestational age at delivery (Table 5B). At 1 week of age we found no significant associations between the NNNS scores and RBC LC-PUFA, but infant RBC-AA was found to be positively associated with regulation 1 month after delivery (Table 5B). This association was also significant after adjusting for AA status at 1 week (p=0.028). Adjusting for regulation at 1 week of age had no impact on this association. Infant RBC-DHA at 1 month or the change over time in RBC-DHA was not associated with any of the NNNS scores at 1 month. The changes in NNNS scores between the two examinations and the corresponding changes in RBC-AA and DHA were not associated.

LC-PUFAs and BSID-III

T-tests revealed a gender difference in the score on the cognitive scale with girls having 14% higher scores than boys (p=0.045). No other gender differences were found in the BSID-III scores. The correlations between the breast-milk sample LC-PUFA means and infant RBC LC-PUFA at 1 month of age versus BSID-III outcomes were generally not significant, except for breast-milk AA, which correlated positively with the cognitive scale score (p=0.002, r=0.46). The mean AA for all breast-milk samples was also associated with the cognitive score in models adjusted for gender (B=8.69±3.08, p=0.007). Furthermore, RBC-AA was associated with the receptive language and fine motor scale score in univariate adjusted GLM models, but in this it was an inverse association (Table 6). Inclusion of infant NNNS scores at 1 week as covariates did not change the associations between RBC LC-PUFA and BSID-III.
PUFA and BSID-III scores. These models did not show any significant associations between the NNNS scores at 1 week and the BSID-III scores at 1 year, except for QoM which was associated with expressive language (B=1.35±0.63, p=0.040).

**Discussion**

Our most important findings were that AA and DHA in the breast-milk and infant RBC samples were significantly associated and that both early and later neurodevelopment of late-preterm infants was not related to RBC-DHA, but related to infant RBC-AA status. The fatty acid composition of the breast-milk samples demonstrated a considerable individual variation. Compared to the present study, previous Danish and Dutch studies [26,34] found lower breast-milk levels of AA and DHA, and similar levels of LA in mothers of term infants with low-average fish intake. These studies also reported a reduction in the breast-milk content of AA and DHA from delivery.
to later stages of breastfeeding as well as an increase in the LA content [26,34]. Two Swedish studies on moderately-late preterm infants have also shown lower breast-milk levels of all three PUFAs [5,34], but a Spanish study found a considerably higher breast-milk levels of LA and AA levels, which were similar to the ones we observed [13]. The Spanish study did not include any breast-milk samples from late-stage lactation, but found that the DHA level increased over the first few weeks of lactation from initially 0.1% to 0.5% of the fatty acids [13].

The individual levels of both LA, AA and DHA in the RBC from the infants in the present study tended to be higher than that reported in more preterm populations [35-37], but resembled reported levels in RBC from both Danish [25] and American term infants [38,39].

Table 4: Developmental scores on Bayley Scales of Infant Development-III at 1 year corrected-age.

<table>
<thead>
<tr>
<th>Sampling time</th>
<th>LA</th>
<th>AA</th>
<th>DHA</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 week</td>
<td>NS</td>
<td>p=0.04</td>
<td>r=0.33</td>
</tr>
<tr>
<td>1 month</td>
<td>p=0.00</td>
<td>r=0.51</td>
<td>p=0.00</td>
</tr>
<tr>
<td>3 months&lt;sup&gt;1&lt;/sup&gt;</td>
<td>NS</td>
<td>p=0.03</td>
<td>r=0.44</td>
</tr>
</tbody>
</table>

Data are given as Spearman’s correlation coefficient (r) and p-value. NS=non-significant. LA=linoleic acid, AA=arachidonic acid. DHA=docosahexaenoic acid. RBC=erythrocyte.

<sup>1</sup>Corrected age

Table 3: Correlations between fatty acids in breast-milk and infant erythrocytes (RBC).

<table>
<thead>
<tr>
<th></th>
<th>Male (n=30)</th>
<th>Female (n=18)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>Range</td>
</tr>
<tr>
<td>Cognitive scale score</td>
<td>12.0 ± 3.1</td>
<td>5 - 18</td>
</tr>
<tr>
<td>Receptive scale score</td>
<td>8.9 ± 2.4</td>
<td>5 - 15</td>
</tr>
<tr>
<td>Expressive scale score</td>
<td>10.5 ± 2.0</td>
<td>7 - 15</td>
</tr>
<tr>
<td>Fine motor scale score</td>
<td>11.5 ± 2.4</td>
<td>8 - 16</td>
</tr>
<tr>
<td>Gross motor scale score</td>
<td>8.7 ± 2.0</td>
<td>4 - 13</td>
</tr>
</tbody>
</table>

Data are given as mean ± range and (min-max).

A few previous studies have found that LC-PUFA composition of breast-milk/formula, maternal RBC and umbilical cord blood was associated with the early QoM in term infants [18–20] as well as in preterm infants (born in gestational week 24-36) [17,35], but the investigated LC-PUFA indices and ratios vary and the results are not very consistent. None of these studies have tested the infants as shortly after delivery as in the present study. However, we found no associations between LC-PUFA and the NNNS scores at 1 week. Our results did however show a positive association between infant RBC-AA (both at 1 week and 1 month) and the NNNS score for regulation at 1 month of age. RBC-AA at 1 month of age was also found to be negatively associated with BSID-III receptive language and fine motor abilities at 1 year corrected age, but no associations were observed for DHA. The lack of a significant associations between breast-milk or RBC-DHA and developmental outcome are in line with previous observations [40–42], although some studies have demonstrated beneficial effects of DHA on psychomotor development in both term and preterm infants [35,43–45]. A recent Swedish study found significant correlations between breast-milk AA and developmental outcomes at 40 weeks gestational age in preterm infants (born in gestational week 24-36) using the Brazelton Neonatal Behavioral Assessment Scale, which is very similar to the NNNS [16]. Another Swedish cohort study [34] of moderately preterm infants found positive associations between both AA and DHA in infant blood and global developmental outcomes at 18 months of age, but negative associations with n-6 PUFA and the n-6/n-3 PUFA ratio. The observed association between AA and language skills is in accordance with some previous studies of term infant showing negative associations between LC-PUFA intake or status and language development [26,46], but other studies found no effect [47,48]. The previous studies have primarily focused on the effect of supplementation with fish oil, DHA or DHA+AA. Recent large randomized controlled trials have shown a negative effect of maternal DHA supplementation in pregnancy on the language development in girls born at term [48], but a positive effect of post-delivery DHA

Table 5: Quality of movement (QoM), regulation and arousal at 1 week to 1 month after delivery (A) and their association with infant erythrocyte arachidonic acid (AA) and docosahexaenoic acid (DHA) (B)

<table>
<thead>
<tr>
<th></th>
<th>1 week</th>
<th>1 month</th>
<th>Change over time (A NNNS)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>Range</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>QoM (3.81±0.78)</td>
<td>4.51 ± 0.41</td>
<td>3.00–5.17</td>
<td>4.48 ± 0.47</td>
</tr>
<tr>
<td>Regulation (5.0 ± 0.82)</td>
<td>5.65 ± 0.76</td>
<td>4.00–7.67</td>
<td>5.91 ± 0.83</td>
</tr>
<tr>
<td>Arousal (4.16 ± 0.81)</td>
<td>3.46 ± 0.71</td>
<td>2.00–5.00</td>
<td>4.02 ± 0.68</td>
</tr>
<tr>
<td><strong>B</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>QoM</td>
<td>0.00 ± 0.04</td>
<td>0.95</td>
<td>50</td>
</tr>
<tr>
<td>Regulation</td>
<td>0.00 ± 0.07</td>
<td>0.99</td>
<td>50</td>
</tr>
<tr>
<td>Arousal</td>
<td>-0.06 ± 0.07</td>
<td>0.42</td>
<td>50</td>
</tr>
<tr>
<td><strong>DHA</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>QoM</td>
<td>0.12 ± 0.06</td>
<td>0.06</td>
<td>50</td>
</tr>
<tr>
<td>Regulation</td>
<td>0.04 ± 0.12</td>
<td>0.74</td>
<td>50</td>
</tr>
<tr>
<td>Arousal</td>
<td>0.10 ± 0.12</td>
<td>0.40</td>
<td>50</td>
</tr>
</tbody>
</table>

Data are given as mean ± SD and range, or parameter estimates (B) ± SE, p-values and n. All analyses are adjusted for gender and gestational age at delivery.
supplementation in preterm girls and no effect in boys [48,49].

The individual variation in DHA status at a given time was substantial, but it may take a considerably larger population to detect a potential effect of DHA on development. The lack of an association between DHA and development could also be due to a lack of n-3 PUFA deficiency in the infants in the study. DHA and AA are usually measured as mass-% and “compete” with each other as well as other PUFA with respect to RBC-incorporation, it is therefore possible that the seemingly negative effect of AA to some extent is caused by reduced presence of other PUFAs. The functional domains affected by RBC-AAs in this study (regulation, receptive language and fine motor skills) share a relation with the periventricular midbrain structures and the cerebellum, which are where the accretion of LC-PUFA mainly occur in late gestation. DHA accumulates in the brain at a later stage than AA [4] and the impact of early sub-optimal DHA supply may only be found in later stages of neurodevelopment.

As described above, infant RBC-AA was linked to neurodevelopmental outcomes in both early and later infancy, but in an inconsistent way. We are not able to say if the early benefit and later negative effect in this study are linked or if they are due to chance, as we were unable to show an association between early neurodevelopmental maturity and psychomotor scores at 1 year, except for an association between QoM at 1 month of age and the expressive language score of BSID-III. It is possible that a larger sample size would have enabled us to demonstrate more associations between early neuromaturation and later psychomotor development.

Neurodevelopment both early in infancy and at 1 year was assessed with clinically validated tests. Our three selected NNNS summary scores (QoM, regulation and arousal) were related to what we wanted to examine (motor control, stimulus tolerance and level of alertness) and gave reliable scores (i.e. were available in most of the infants, normally distributed and internally correlated). These functional domains also relate to brain areas associated with perinatal LC-PUFA accretion. QoM in early infancy has been addressed quite often in previous studies and is used as a measure of motor maturity in several other developmental tests (e.g., Prechtl, ENIGMA or Peabody). Regulation is a measure of the infants ability to tolerate stimulation and is closely related to habituation, which is often used as a measure of the ability to inhibit responses to a continued stimulus (tactile, visual or auditory). However, the evaluation of infant habituation in the NNNS test requires that the test is started with a sleeping infant, and in an outpatient setting such as ours, this is not always possible. BSID-III is a global developmental test, designed to contrast developmental delay and normal psychomotor development. It is considered the golden standard for developmental testing of infants and is widely used in studies of LC-PUFA influences on early childhood development, but some researchers have pointed out that it may not be ideal for detecting minor variations within normality [47,50-54].

The main limitations of the present study is the small sample size. We planned for a considerably larger study, but were not able to include more participants within the given time frame. This was mainly explained by a 40% drop in the local preterm infant delivery rate [55] due to a temporary discontinuation of a longstanding government subsidy for infertility treatment. Furthermore, the exposures (2) and outcomes (8) were tested multiple times against each other. Formal Bonferroni correction (p<0.003) would yield all of our tests insignificant, but the Bonferroni method is known for its very conservative approach and we found it un-suited for our explorative purpose [56]. We realize that much caution must be applied in the interpretation of our results, but they can be used for the planning future studies of LC-PUFA and infancy development. Depending on the desired outcome, and given the parameter estimates in this study, the required sample size in such studies would have to be considerably larger, e.g. 111 and 214 for QoM and arousal at 1 month of age, respectively, and 345 for BSID-III at 1 year.

Due to the observational nature of the study we cannot exclude residual confounding although we did adjust for various maternal variables, but the possibility of other confounders such as maternal stimulation of the infant or genetic influences remain. Furthermore, only 62% (n=33) were able to exclusively breastfeed and there was a highly significant difference in the frequency of maternal smokers between those still breastfeeding at 3 months corrected age and those with a short breastfeeding duration. This is a potential bias in our study, as it resulted in a higher representation of breast-milk samples from women of higher socioeconomic status. The associations between the RBC samples and the developmental outcomes were, however, not affected by maternal smoking. There was a slight difference in maternal age between the participants and the non-participants, but this difference is probably not a clinically relevant factor. However, we did find a minor, but significant, difference in the occurrence of maternal psychiatric disorder between the infants lost to follow-up and those remaining in the study, but this were caused by only one twin mother. Maternal psychiatric disorder was a statistically significant covariate in the associations between infant LC-PUFA status and receptive language BSID-III score, but because of the small sample size, the impact of this difference is uncertain. The overall attendance at outpatient controls and follow-up was good and the consistency of our data is good. The internal validity of our study is also good as all examinations were performed by one investigator and all samples handled by one lab technician. Furthermore, we did not only rely on breast-milk samples, but also analyzed infant RBC fatty acid composition, which gives a more stable measure of LC-PUFA accretion.

Conclusion

In this relatively small cohort, late-preterm breast-milk LC-PUFA content was high compared to other preterm studies. The infants were primarily healthy with appropriate weight for gestational age and only minor developmental variations, but certain aspects of early neuromaturation and development at 1 year corrected age was associated in an inconsistent way with AA in breast-milk and infant RBC in the perinatal period. Our results must be approached with caution, but they were further testing with precise tests of sensory integrity in larger cohorts to properly investigate effects of perinatal LC-PUFA-status on early neurodevelopment in healthy late-preterm infants.

<table>
<thead>
<tr>
<th>BSID-III</th>
<th>Infant erythrocyte (n=44)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>AA</td>
</tr>
<tr>
<td>Cognitive scale score</td>
<td>B=-0.28 ± 0.28, p=0.34†</td>
</tr>
<tr>
<td>Receptive scale score</td>
<td>B=0.53 ± 0.26, p=0.05§</td>
</tr>
<tr>
<td>Expressive scale score</td>
<td>B=-0.14 ± 0.16, p=0.40¶</td>
</tr>
<tr>
<td>Fine motor scale score</td>
<td>B=-0.70 ± 0.30, p=0.03¶</td>
</tr>
<tr>
<td>Gross motor scale score</td>
<td>B=0.19 ± 0.22, p=0.38</td>
</tr>
</tbody>
</table>

Data are given as parameter estimates (B ± SD) and p-value (p) from multiple regression analysis adjusted for 1 Gender; 2 Gender, corrected-age; 3 Gender, maternal psychiatric disorder; 4 Multiple birth; and 5 Multiple birth, corrected age at 1 year corrected age measured with Bayley Scales of Infant Development-III.

Table 6: Univariate associations between infant erythrocyte arachidonic acid (AA) and docosahexaenoic acid (DHA) at 1 month of age and developmental outcomes.
Conflict of Interest
None

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References
2. P. Guesnet and JM Alessandri (2011) "Docosahexaenoic acid (DHA) and the developing central nervous system (CNS) - Implications for dietary recommendations. Biochimie 93:7-12.


42. Makrides M (2013) DHA supplementation during the perinatal period and neurodevelopment: Do some babies benefit more than others? Prostaglandins Leukot Essent Fatty Acids 88: 87-90.


