



Københavns Universitet

Micronutrient-fortified rice can increase hookworm infection risk

de Gier, Brechje; Campos Ponce, Maiza; Perignon, Marlene; Fiorentino, Marion; Khov, Kuong; Chamnan, Chhoun; de Boer, Michiel R; Parker, Megan E; Burja, Kurt; Dijkhuizen, Marjoleine Amma; Berger, Jacques; Polman, Katja; Wieringa, Frank T

Published in:
P L o S One

DOI:
[10.1371/journal.pone.0145351](https://doi.org/10.1371/journal.pone.0145351)

Publication date:
2016

Document Version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
de Gier, B., Campos Ponce, M., Perignon, M., Fiorentino, M., Khov, K., Chamnan, C., ... Wieringa, F. T. (2016). Micronutrient-fortified rice can increase hookworm infection risk: A cluster randomized trial. P L o S One, 11(1), [e0145351]. <https://doi.org/10.1371/journal.pone.0145351>

RESEARCH ARTICLE

Micronutrient-Fortified Rice Can Increase Hookworm Infection Risk: A Cluster Randomized Trial

Brechje de Gier¹, Maiza Campos Ponce², Marlene Perignon³, Marion Fiorentino³, Kuong Khov⁴, Chhoun Chamnan⁴, Michiel R. de Boer⁵, Megan E. Parker⁶, Kurt Burja⁷, Marjoleine A. Dijkhuizen⁸, Jacques Berger³, Katja Polman^{2,9}, Frank T. Wieringa^{3*}

1 Section Health and Life Sciences, Athena Institute, Faculty of Earth and Life Sciences, VU University Amsterdam, Amsterdam, The Netherlands, **2** Section Infectious Diseases, department of Health Sciences, Faculty of Earth and Life Sciences, VU University Amsterdam, Amsterdam, The Netherlands, **3** UMR-204, NutriPass IRD-UM-SupAgro, Institut de Recherche pour le Développement, Montpellier, France, **4** Department of Fisheries Post-Harvest Technologies and Quality control, Fisheries Administration, Phnom Penh, Cambodia, **5** Section Methodology and Applied Biostatistics, Department of Health Sciences, Faculty of Earth and Life Sciences, VU University Amsterdam, Amsterdam, The Netherlands, **6** PATH, Seattle, Washington, United States of America, **7** United Nations World Food Programme, Phnom Penh, Kingdom of Cambodia, **8** Department of Nutrition, Exercise and Sports, Copenhagen University, Copenhagen, Denmark, **9** Department of Biomedical Sciences, Institute for Tropical Medicine, Antwerp, Belgium

* franck.wieringa@ird.fr



OPEN ACCESS

Citation: de Gier B, Campos Ponce M, Perignon M, Fiorentino M, Khov K, Chamnan C, et al. (2016) Micronutrient-Fortified Rice Can Increase Hookworm Infection Risk: A Cluster Randomized Trial. PLoS ONE 11(1): e0145351. doi:10.1371/journal.pone.0145351

Editor: David Joseph Diemert, The George Washington University School of Medicine and Health Sciences, UNITED STATES

Received: May 5, 2015

Accepted: December 1, 2015

Published: January 6, 2016

Copyright: © 2016 de Gier et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data have been reported in the paper.

Funding: The research described in this paper received funding from United States Department of Agriculture/FAS through a grant (FFE-442-2012/038-00, 10.608) to PATH and internal funding from WFP (through the WFP/DSM partnership) and IRD. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

Background

Fortification of staple foods is considered an effective and safe strategy to combat micronutrient deficiencies, thereby improving health. While improving micronutrient status might be expected to have positive effects on immunity, some studies have reported increases in infections or inflammation after iron supplementation.

Objective

To study effects of micronutrient-fortified rice on hookworm infection in Cambodian schoolchildren.

Methods

A double-blinded, cluster-randomized trial was conducted in 16 Cambodian primary schools partaking in the World Food Program school meal program. Three types of multi-micronutrient fortified rice were tested against placebo rice within the school meal program: UltraRice_original, UltraRice_improved and NutriRice. Four schools were randomly assigned to each study group (placebo n = 492, UltraRice_original n = 479, UltraRice_improved n = 500, NutriRice n = 506). Intestinal parasite infection was measured in fecal samples by Kato-Katz method at baseline and after three and seven months. In a subgroup (N = 330), fecal calprotectin was measured by ELISA as a marker for intestinal inflammation.

Competing Interests: The authors have declared that no competing interests exist.

Results

Baseline prevalence of hookworm infection was 18.6%, but differed considerably among schools (range 0%–48.1%). Micronutrient-fortified rice significantly increased risk of new hookworm infection. This effect was modified by baseline hookworm prevalence at the school; hookworm infection risk was increased by all three types of fortified rice in schools where baseline prevalence was high (>15%), and only by UltraRice_original in schools with low baseline prevalence. Neither hookworm infection nor fortified rice was related to fecal calprotectin.

Conclusions

Consumption of rice fortified with micronutrients can increase hookworm prevalence, especially in environments with high infection pressure. When considering fortification of staple foods, a careful risk-benefit analysis is warranted, taking into account severity of micronutrient deficiencies and local prevalence of parasitic infections.

Trial Registration

ClinicalTrials.gov [NCT01706419](https://clinicaltrials.gov/ct2/show/study/NCT01706419)

Introduction

Micronutrient deficiencies form a large public health problem worldwide, especially in tropical regions[1]. Children are particularly vulnerable, due to their specific nutritional needs for growth and development. Approximately 50% of all child mortality has been attributed to malnutrition, including deficiencies of iron, vitamin A and zinc[2, 3]. Aside from mortality, micronutrient deficiencies affect growth and cognitive development[4, 5]. Micronutrient deficiencies are frequently combated by micronutrient supplementation or fortification of staple foods [1]. Indeed, the Copenhagen Consensus ranks food fortification as one of the most cost-effective tools to combat malnutrition[6].

The world regions where micronutrient deficiencies are the most common are also often plagued by high prevalence of helminth infections. Associations between micronutrients and helminth infections have been reported, although many questions remain unanswered[7]. Micronutrient deficiencies can increase susceptibility to infection, but infections can also alter the intestinal mucosa, leading to reduced absorption of nutrients. This phenomenon is being increasingly recognized as environmental enteropathy[8]. On the other hand, micronutrient fortification might even increase infection risk or persistence. This phenomenon has been described for iron supplementation and several pathogens[9]. The debate surrounding this conundrum has been fueled by a trial in Pemba, Tanzania, in which mortality for malaria and other infections was higher in children who were given iron and folate supplements[10]. Since then, systematic reviews have been performed but have not found a significantly increased infection risk after iron or multi-micronutrient supplementation[7, 11].

Aside from infections, the intestinal environment might be altered in other ways by micronutrient supplementation. In 2010, Zimmermann et al found increases in intestinal inflammation and enterobacteria after iron supplementation[12]. These findings raise questions about the effects of micronutrient supplementation or fortification on the intestinal environment and immunity.

Despite considerable improvements in health and nutrition since the 1990's, Cambodian children are at high risk of stunting, wasting and micronutrient deficiencies[13–15]. A 2010 national survey found a prevalence of stunting in children <5 of almost 40%[15]. The

prevalence of helminths, mainly of hookworm, is also considerable in Cambodia[16]. The FORISCA UltraRice+NutriRice project was a large scale cluster-randomized, double-blinded, placebo-controlled trial, conducted in 16 primary schools, all situated in one province in Cambodia. The project aimed to quantify the impact of multi-micronutrient fortified rice, which was distributed through the World Food Program (WFP) school meal program as a single meal per day, on micronutrient status, health and cognition of Cambodian schoolchildren. As uncertainly exists on the most optimal combination of micronutrients to be added to rice, 3 different types of fortified rice, produced using different methods and with different micronutrient composition, were studied. Secondary outcomes included helminth infection, anthropometry and intestinal inflammation. Here we report on effects of the introduction of fortified rice on hookworm infection and local intestinal inflammation.

Methods

Study design and population

In a double-blinded, cluster-randomized, placebo-controlled trial, three different types of multi-micronutrient fortified rice were introduced through the World Food Program (WFP) School Meal program in Cambodia. The clusters were 16 primary schools in rural Kampong Speu province, of which four were randomly selected for each study group. Schools were eligible if they participated in the WFP school meal program and all children were served breakfast daily. In total 18 schools were eligible, two were excluded because of the number of school children (one school had double the number of school children ($N > 1200$) as the other schools, and one school had < 100 school children, whereas for biochemical determination of micronutrient status a minimum of 125 school children was required per school). A cluster-randomization was chosen because the schools had one kitchen each, and separate preparations of school meals were not feasible. The study was powered for its primary outcomes (micronutrient status), which are not reported here. The trial took place from November 2012 to June 2013. The clusters were 16 primary schools in rural Kampong Speu province, of which four were randomly selected for each study group. Calprotectin was measured in a subsample, from two schools from the placebo, UltraRice_original and UltraRice_improved study groups due to financial restraints. Written informed consent of at least one parent was obtained prior to the study. Ethical approval was obtained from the Cambodian Ministry of Health, Education and Planning and the Ethical Review board of PATH, USA. This study population is further described by Perignon et al[17].

Intervention

The 3 types of fortified rice differed in micronutrient compositions (Table 1), as well as production procedures. NutriRice was produced by hot extrusion by Buhler Food, Wuxi, China. Both types of UltraRice were custom-made for the project, with UltraRice_original produced using cold extrusion techniques by Maple Grove Gluten-free Foods, Ltd, California, USA and UltraRice_improved by the Food Technology department of Kansas State University, USA. Children received one type of fortified rice or placebo (unfortified white rice) six days per week for six months. Aside from rice, the school meals consisted of canned fish, vitamin A+D fortified vegetable oil, yellow split peas and iodized salt. After baseline data collection, all children received a single dose of 500 mg mebendazole.

Measurements

The primary outcome for the current paper is hookworm infection, which was the main intestinal parasite found in this population. Stool samples were collected by providing school children

Table 1. Micronutrient composition of the three types of fortified rice. N.I. = not included in premix

Micronutrient	Target value(mg)	UltraRice _original(mg)	UltraRice _improved(mg)	NutriRice (mg)
Vitamin A (retinol)	0.3	N.I.	0.64	0.29
Iron	7.26	10.67	7.55	7.46
Zinc	3.5	3.0	2.0	3.7
Vitamin B1 (Thiamin)	0.6	1.1	1.4	0.7
Vitamin B3 (NE)	8	N.I.	12	8
Vitamin B6	0.65	N.I.	N.I.	0.92
Folate (DFE)	0.2	0.2	0.3	0.1
Vitamin B12	0.001	N.I.	0.004	0.001

doi:10.1371/journal.pone.0145351.t001

with a sample collection container and requesting them to return the next day to school with a fresh stool sample. Sample containers were collected in the morning, stored in a coolbox with blue ice (4°C) and brought to the National Center for Parasitology, Entomology and Malaria control (CNM) in Phnom Penh in the afternoon. Samples were analyzed by Kato-Katz technique at baseline (before treatment), three months and seven months (end of the intervention) to determine helminth infection[18]. Parasite diagnosis was performed by the National Center for Parasitology, Entomology and Malaria control (CNM), Phnom Penh, Cambodia, and recorded as eggs per gram of feces. No distinction was made between hookworm species. For a subgroup of 330 children, at baseline and after seven months stool samples were frozen (-20°C) and sent to the Institute for Tropical Medicine in Antwerp, Belgium where our secondary outcome calprotectin was measured by ELISA (Calpro AS, Norway), according to the manufacturer’s instructions, with 10% of these samples measured in duplicate for quality control. Due to funding restraints, fecal calprotectin was measured only in a subsample of stool samples collected after seven months from UltraRice_original, UltraRice_improved and placebo groups. Sex and age of the children were obtained by interviews and verified by school records and birth certificates. All measurements were at the individual level.

Randomization and masking

Three different randomizations, combining different schools to one intervention, were separately generated based on a list number of children per school by iteration to fit the predefined criteria of group size (within 10% of the mean). A researcher not involved in the field work (MAD) blindly picked one of the three randomizations, and allocated each group of schools to an intervention arm. To further assure blinding, each intervention arm of 4 schools was split into two groups of two schools, each given a letter code (A–H). The entire research team and all participants and caregivers were blinded to the allocation. The code was only known to one person with WFP, responsible to allocate the correct type of rice to the right school. The rice packaging was coded with the letter allocated during the randomization (A–H) and did not contain the name of the rice type.

Statistical analysis

Analyses were done using SPSS software version 22 (IBM, NY, USA). Crude percentage points differences in hookworm prevalence (with corresponding confidence intervals for proportions) between the intervention and placebo groups were calculated. Effects of the interventions on hookworm infection risk were analyzed using generalized linear mixed models. A logistic mixed model with hookworm infection as the binary outcome at three and seven months and random intercepts for school and child ID was used to account for repeated measurements

within subjects and clustering of subjects within schools. Covariates were sex, age in quartiles and baseline hookworm prevalence at the school. In the analysis we focused on the effect of the intervention on hookworm infection in children who were uninfected at baseline, in order to estimate new infection rate. The fecal calprotectin data were skewed, which could not be corrected by logarithmic transformation. Therefore, fecal calprotectin was analyzed using mixed model analysis with high fecal calprotectin (>50 mg/kg) at seven months as binary outcome and a random intercept for school clustering. Covariates were sex, age in quartiles and high fecal calprotectin at baseline (binary) [19]. Effect modification was examined by introducing interaction terms into the model, when these showed significant effects ($p < 0.05$), we stratified the analysis.

Results

Rice kernels were analyzed for micronutrient composition by Silliker (Markham, Ontario, Canada) (for both UltraRice kernels) and Buhler (NutriRice kernels; Table 1). A flow chart of all study participants is shown in Fig 1. At baseline, 3 children were excluded due to severe anemia (and received multiple micronutrient supplements for 2 months). Stool samples were obtained from 1393 children at baseline (70.5%, Table 2). After three and seven months, fecal samples were obtained from 1256 (63.5%) and 1257 (63.6%) children, respectively.

Overall prevalence of hookworm was 18.6% (baseline), 25.4% (three months) and 28.7% (seven months). Hookworm infections were mostly of light intensity (<2000 eggs per gram feces) and no heavy infections (>4000 egg per gram) were found. Baseline prevalence of hookworm in the 16 schools ranged from 0% to 48.1%. Two thirds of the infected children at baseline were also infected after three months. We cannot ascertain whether these are all re-infections or if the anthelmintic treatment provided after baseline had been ineffective, as no stool samples were obtained shortly after treatment. Therefore, to determine whether hookworm infection rate was increased after the introduction of fortified rice, we focused our further analysis on the children who were uninfected at baseline. Fig 2 shows the prevalence of hookworm infections over the course of the study in all included children (A) and baseline uninfected children (B) per intervention group. S1 Fig shows the prevalence of hookworm infection at baseline, after 3 and 7 months for each school, by study group.

New hookworm infection rates were significantly higher in schools receiving any type of fortified rice compared to placebo. The increase in hookworm infection was significant for both types of UltraRice (Table 3, Fig 2B). Baseline prevalence of hookworm at the school level was a significant modifier of the effects of fortified rice on hookworm infection risk. Therefore we stratified for baseline hookworm prevalence (below and above the median of 15%, Table 4). In schools with baseline hookworm prevalence below 15%, only UltraRice_original increased infection risk. In schools with baseline prevalence above 15%, all three types of fortified rice significantly increased the risk of infection compared to placebo.

Fecal calprotectin concentrations had increased after seven months in all groups (Table 5). Neither type of UltraRice had a significant effect on fecal calprotectin concentration compared to placebo. Calprotectin and hookworm infection were not associated at either baseline or at seven months.

Discussion

To our knowledge, this is the first study to show a significantly increased risk of hookworm infection in children receiving multi-micronutrient fortified rice. Negative effects of iron supplements on hookworm infection prevalence have been reported in adults, although this appeared to be a transient effect [20]. In our study, the effect of fortified rice on hookworm

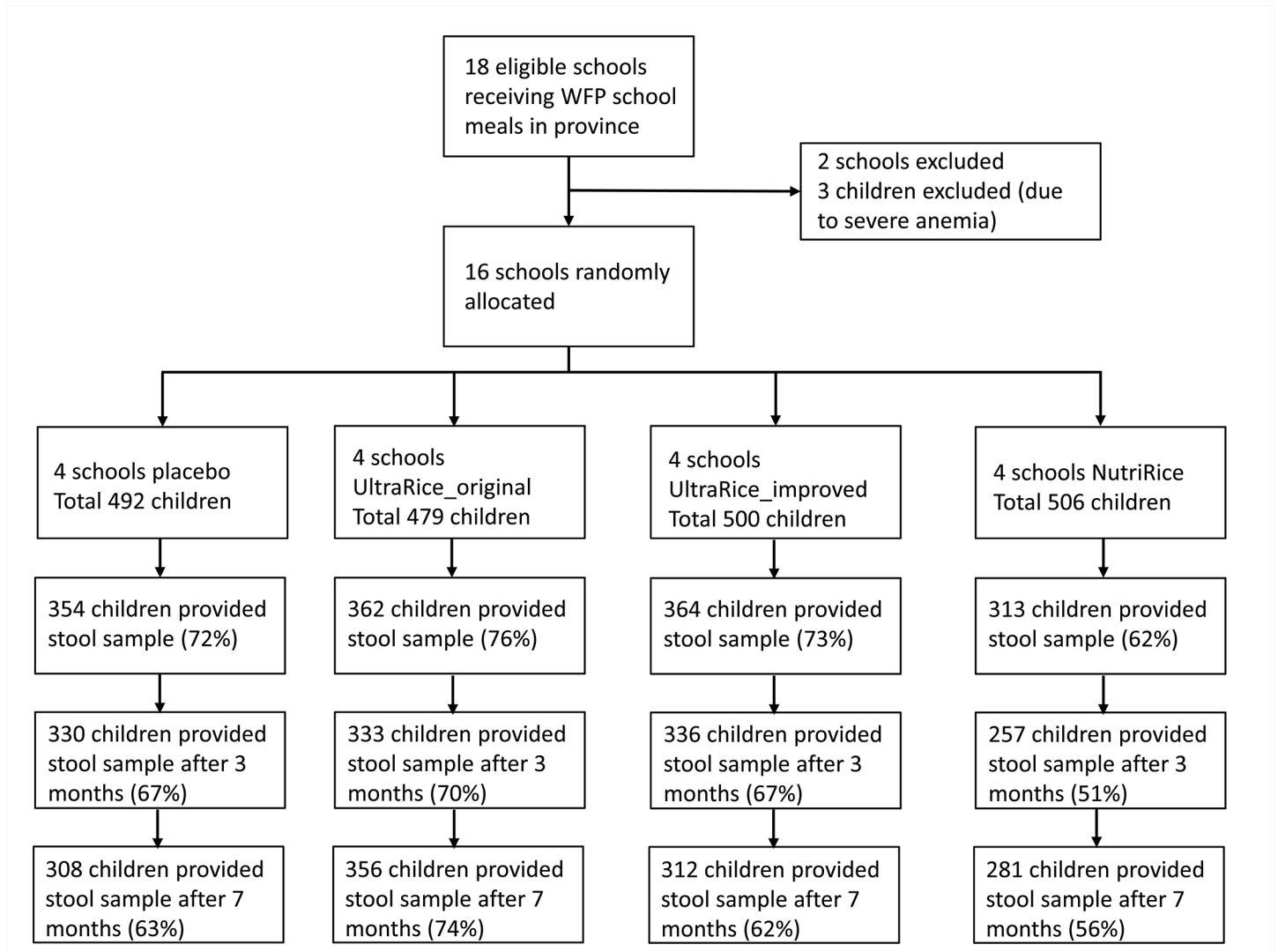


Fig 1. Flow chart of the study.

doi:10.1371/journal.pone.0145351.g001

Table 2. Population characteristics at baseline.

	Placebo	UltraRice_original	UltraRice_improved	Nutrice
N (male %)	492 (49.9%)	479 (49.6%)	500 (49.8%)	506 (50.8%)
Age (mean ± SD)	9.58 ± 2.27	9.61 ± 2.19	9.64 ± 2.22	9.71 ± 2.42
Hookworm infection n/N (%)	74/354 (20.9%)	82/362 (22.7%)	67/364 (18.4%)	36/316 (11.5%)
Range of baseline hookworm prevalence at schools	8.0–37.6%	0–48.1%	5.1–33.3%	4.8–26.9%
Hookworm infection intensity n				
-Light	70	81	65	36
-Moderate	4	1	2	0
-Heavy	0	0	0	0
Calprotectin N	124	117	123	Na
Calprotectin median (IQR)	22.7 (21.2 to 26.3)	22.7 (20.2 to 26.9)	21.7 (20.3 to 26.3)	Na
Calprotectin high n (>50mg/kg)	13 (10.5%)	7 (6%)	13 (10.6%)	Na

doi:10.1371/journal.pone.0145351.t002

prevalence was modified by the baseline prevalence of hookworm at the schools, indicating that infection pressure is of importance.

The strong modifying effect that school hookworm prevalence had on the hookworm infection risk effects of fortified rice warrants caution when implementing micronutrient

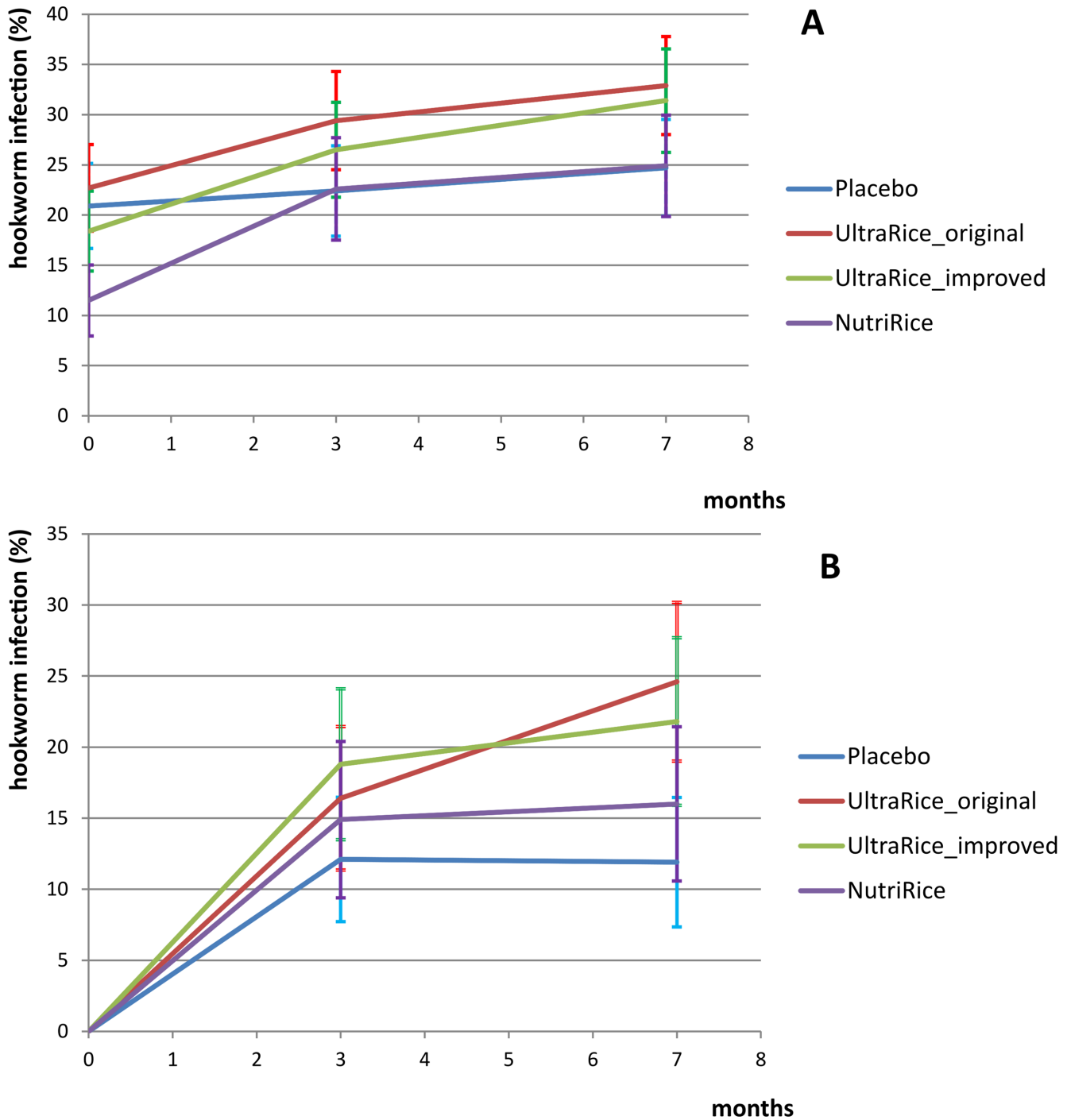


Fig 2. Effect of micronutrient-fortified rice consumption or placebo on hookworm infection. Hookworm infection prevalence in all children (A, N = 1393) and baseline uninfected children (B, N = 1134) per study group, during the course of the intervention, with error bars depicting crude 95% confidence intervals for the proportion infected.

doi:10.1371/journal.pone.0145351.g002

Table 3. Longitudinal effects of micronutrient fortified rice on hookworm infection risk of baseline uninfected children.

Hookworm infection: baseline uninfected children							
	Baseline n/N (%)	3 months n/N (%)	7 months n/N (%)	crude % point risk difference with placebo (95% CI)	aOR ¹ (N = 982)	95% CI	P value
Placebo	0/280 (0.0)	26/214 (12.1)	23/194 (11.9)		1 (ref)		
Any fortified rice	0/854 (0.0)	97/576 (16.8)	125/591 (21.2)	+ 9.3% (2.9 to 14.7)	1.86	1.17 to 2.95	0.009
- UltraRice_ original	0/280 (0.0)	34/207 (16.4)	56/228 (24.6)	+ 12.7% (5.3 to 19.8)	1.89	1.06 to 3.37	0.032
- UltraRice_improved	0/297 (0.0)	39/208 (18.8)	41/188 (21.8)	+ 10.0% (2.5 to 17.4)	1.91	1.12 to 3.26	0.018
- NutriRice	0/277 (0.0)	24/161 (14.9)	28/175 (16.0)	+ 4.1% (-2.9 to 11.4)	1.74	0.63 to 4.81	0.284

¹ From mixed model analysis, adjusted for sex, age (in quartiles) and baseline prevalence of hookworm at the school.

doi:10.1371/journal.pone.0145351.t003

supplementation strategies in endemic areas. Even though all schools were in the same province, we found large differences in baseline hookworm prevalence across schools. Our results show that even within the same province, large regional differences can exist in the health effects of consumption of multi-micronutrient fortified rice by school children. Public health policies aimed at improving child micronutrient status should take hookworm infection risk into account, as hookworms are known to induce iron deficiency through blood loss [21]. Our results suggest that aside from anthelmintic treatment at schools, the abundance of hookworm eggs and larvae in the environment needs to be reduced to safeguard the children from (re)infection.

The increase in hookworm prevalence in all groups was surprising, given the anthelmintic treatment that was provided after baseline measurements. However, mebendazole has recently been shown to be significantly less effective than albendazole against hookworm in Cambodia [22]. The overall increase of infection might be a seasonal effect. It was also unexpected that 52 children who were infected at three months seemed uninfected at seven months, since no

Table 4. Effect of micronutrient fortified rice on hookworm infection risk of baseline uninfected children, stratified by baseline school hookworm prevalence.

Hookworm infection risk by school prevalence category						
Baseline prevalence < 15%						
	Baseline n/N (%)	3 months n/N (%)	7 months n/N (%)	aOR ¹ (N = 493)	95% CI	P value
Placebo	0/92(0.0)	15/82 (18.3)	4/74 (5.4)	1 (ref)		
Any fortified rice	0/477 (0.0)	35/312 (11.2)	44/311 (14.1)	0.96	0.62 to 1.47	0.840
- UltraRice_ original	0/87(0.0)	13/64 (20.3)	19/69 (27.5)	2.29	2.13 to 2.46	<0.001
- UltraRice_improved	0/162(0.0)	11/116 (9.4)	10/91 (10.9)	0.82	0.49 to 1.37	0.454
- NutriRice	0/228(0.0)	11/132 (8.3)	15/151 (9.9)	0.73	0.53 to 1.03	0.070
Baseline prevalence >15%						
	Baseline n/N (%)	3 months n/N (%)	7 months n/N (%)	aOR ¹ (N = 489)	95% CI	P value
Placebo	0/139(0.0)	11/132 (8.3)	19/120 (15.8)	1 (ref)		
Any fortified rice	0/377 (0.0)	62/264 (23.5)	81/280 (28.9)	2.94	1.82 to 4.77	<0.001
- UltraRice_ original	0/193(0.0)	21/143 (14.7)	37/159 (26.3)	1.82	1.33 to 2.49	<0.001
- UltraRice_improved	0/135(0.0)	28/92 (30.4)	31/97 (32.0)	3.73	2.76 to 5.05	<0.001
- NutriRice	0/49(0.0)	13/29 (44.8)	13/24 (54.2)	8.65	6.59 to 11.34	<0.001

¹From mixed model analysis, adjusted for sex and age (in quartiles).

doi:10.1371/journal.pone.0145351.t004

Table 5. Effects of UltraRice_original and UltraRice_improved on prevalence of elevated fecal calprotectin (>50 mg/kg).

	Fecal calprotectin >50 mg/kg				
	Baseline n/N (%)	7 months n/N (%)	aOR ¹	95% CI	P value
Placebo	13/124 (10.5)	39/101 (38.6)	1 (ref)		
Any fortified rice	20/240 (8.3)	78/243 (32.1)	0.69	0.46 to 1.02	0.065
- UltraRice_original	7/117 (6.0)	39/121 (32.2)	0.74	0.32 to 1.72	0.481
- UltraRice_improved	13/123 (10.6)	39/122 (32.0)	0.66	0.29 to 1.51	0.324

¹ From mixed model analyses, adjusted for sex, age (in quartiles), baseline calprotectin >50 mg/kg yes/no, N = 330

doi:10.1371/journal.pone.0145351.t005

treatment was given at the schools between those time points. We suspect these to be false negatives; the Kato Katz diagnostic method is not very sensitive for low intensity hookworm infections [23]. Treatment received outside of school or study programs could also explain this observation. These 52 children were randomly distributed over all intervention groups, and definition of these 52 cases as ‘positive’ for hookworm at seven months did not change the findings.

The low number of schools per study group is an important limitation of this study, especially because the prevalence at school level was a large effect modifier. Moreover, the unexpected high variability in baseline hookworm infection prevalence creates imbalance between schools and study groups. The estimated odds ratios were exceptionally high for UltraRice_original in the stratum with low baseline prevalence and for NutriRice in the stratum with baseline prevalence above 15%, however for both these estimates only one school was included. We therefore emphasize the need for better-powered trials to confirm our findings. Also, the amounts of rice consumed by the children were not recorded in this study. Because the three types of fortified rice differed on content of several micronutrients, we cannot draw conclusions about causation of the increased hookworm risk by any one nutrient or amount thereof.

We did not find an effect of micronutrient-fortified rice on intestinal inflammation, measured as fecal calprotectin. Increases in fecal calprotectin after iron supplementation or fortification were found in studies in Ivorian children and Kenyan infants, but not in South African children [12, 24, 25]. Hookworm and calprotectin were not associated with each other at either time point. While calprotectin is considered to be a marker for intestinal inflammation in general, it is mainly derived from neutrophils [19]. Despite hookworms causing mucosal damage, a lack of association between hookworm and neutrophil abundance is to be expected as hookworms express a neutrophil inhibiting factor, thereby perhaps dampening intestinal inflammation [26].

A 2013 systematic review on effects of (multi-) micronutrient fortification studies showed positive effects on micronutrient status but acknowledged the paucity of studies with other health outcomes [27]. A recent study from Vietnam found a large reduction in hookworm prevalence after iron and folic acid supplementation [28]. However, this study did not include a control group. A meta-analysis of (multi-) micronutrient supplementation or fortification on helminth infection risk showed a non-significant increased risk after iron supplementation but a protective effect of multi-micronutrient supplementation [7]. However, when we repeated this meta-analysis including only hookworm as outcome, a non-significant increase in hookworm infection risk after multi-micronutrients was observed. In addition to strengthening the evidence for an increase in hookworm infection risk after multi-micronutrient fortification, we here show that local prevalence also plays an important role in this effect.

During systemic inflammatory (acute phase) responses, micronutrients such as iron and zinc are withheld from the human circulation [29]. This is considered a strategy against parasitic organisms who are also in need of these scarce micronutrients, a phenomenon described

as ‘nutritional immunity’ [30]. Within the context of withholding micronutrients from parasites, supplementation of these nutrients could override nutritional immunity and enhance the infection. This is especially the case for enteric parasites: with increasing micronutrient content of the gut, feeding the parasite instead of the host might become a serious risk. However, as hookworms are not known to feed on luminal contents, an increase in micronutrient concentration of mucosal tissue and blood would be needed to actually feed this parasite. Aside from risk of parasitic infection, the increased micronutrient availability in the gut lumen might influence intestinal microflora composition, which can in turn have a wide range of health consequences [12, 31].

Together, our results raise questions about possible negative health outcomes of micronutrient fortification of staple foods in hookworm endemic areas. Special attention might be warranted for so-called ‘home fortification’ of complementary foods with micronutrient powders. A literature search for the effect of home fortification with micronutrient powders on hookworm infection returned no published papers. We believe further research in this area is urgently needed.

The merits of micronutrient repletion should be weighed carefully against its possible risks. This might need to be considered for every region separately, taking into account local infection prevalence, severity of micronutrient deficiencies and other possible factors of influence. Pairing micronutrient supplementation with vigorous efforts to reduce hookworm infection risk, by frequent administration of anthelmintics and sanitation and hygiene interventions may circumvent the increased risk of hookworm infection, however this would need to be addressed by further studies.

Supporting Information

S1 CONSORT Checklist. CONSORT checklist for the reporting of cluster trials.
(DOCX)

S1 Fig. Hookworm infection prevalence at each school, by study group. N is the number of children at the school for which a baseline hookworm diagnosis was available.
(TIF)

S1 Protocol. The original protocol of the FORISCA study.
(PDF)

Acknowledgments

We are thankful to Kim Vereecken and Liliane Mpabanzi (ITM Antwerp) for the calprotectin measurements.

Author Contributions

Conceived and designed the experiments: FTW JB KB MAD M. Parker. Performed the experiments: MF M. Perignon KK CC M. Parker KB. Analyzed the data: BG MCP MRdB KP FTW. Wrote the paper: BG MCP KP FTW.

References

1. Bhutta ZA, Salam RA, Das JK. Meeting the challenges of micronutrient malnutrition in the developing world. *British medical bulletin*. 2013; 106:7–17. doi: [10.1093/bmb/ldt015](https://doi.org/10.1093/bmb/ldt015) PMID: [23645843](https://pubmed.ncbi.nlm.nih.gov/23645843/)
2. Caulfield LE, de Onis M, Blossner M, Black RE. Undernutrition as an underlying cause of child deaths associated with diarrhea, pneumonia, malaria, and measles. *The American journal of clinical nutrition*. 2004; 80(1):193–8. PMID: [15213048](https://pubmed.ncbi.nlm.nih.gov/15213048/)

3. Black RE, Victora CG, Walker SP, Bhutta ZA, Christian P, de Onis M, et al. Maternal and child undernutrition and overweight in low-income and middle-income countries. *Lancet*. 2013; 382(9890):427–51. doi: [10.1016/S0140-6736\(13\)60937-X](https://doi.org/10.1016/S0140-6736(13)60937-X) PMID: [23746772](https://pubmed.ncbi.nlm.nih.gov/23746772/)
4. Walker SP, Wachs TD, Gardner JM, Lozoff B, Wasserman GA, Pollitt E, et al. Child development: risk factors for adverse outcomes in developing countries. *Lancet*. 2007; 369(9556):145–57. doi: [10.1016/S0140-6736\(07\)60076-2](https://doi.org/10.1016/S0140-6736(07)60076-2) PMID: [17223478](https://pubmed.ncbi.nlm.nih.gov/17223478/)
5. Rivera JA, Hotz C, Gonzalez-Cossio T, Neufeld L, Garcia-Guerra A. The effect of micronutrient deficiencies on child growth: a review of results from community-based supplementation trials. *J Nutr*. 2003; 133(11 Suppl 2):4010S–20S. PMID: [14672304](https://pubmed.ncbi.nlm.nih.gov/14672304/)
6. Hoddinott J, Rosegrant M, Torero M. Hunger and malnutrition. In: Lomborg B, editor. *Global Problems, Smart Solutions*. Cambridge, UK: Cambridge University press; 2013.
7. de Gier B, Campos Ponce M, van de Bor M, Doak CM, Polman K. Helminth infections and micronutrients in school-age children: a systematic review and meta-analysis. *The American journal of clinical nutrition*. 2014; 99(6):1499–509. doi: [10.3945/ajcn.113.069955](https://doi.org/10.3945/ajcn.113.069955) PMID: [24740209](https://pubmed.ncbi.nlm.nih.gov/24740209/)
8. Korpe PS, Petri WA Jr. Environmental enteropathy: critical implications of a poorly understood condition. *Trends in molecular medicine*. 2012; 18(6):328–36. doi: [10.1016/j.molmed.2012.04.007](https://doi.org/10.1016/j.molmed.2012.04.007) PMID: [22633998](https://pubmed.ncbi.nlm.nih.gov/22633998/);
9. Oppenheimer SJ. Iron and its relation to immunity and infectious disease. *The Journal of nutrition*. 2001; 131(2S-2):616S–33S; discussion 33S-35S. PMID: [11160594](https://pubmed.ncbi.nlm.nih.gov/11160594/)
10. Sazawal S, Black RE, Ramsan M, Chwaya HM, Stoltzfus RJ, Dutta A, et al. Effects of routine prophylactic supplementation with iron and folic acid on admission to hospital and mortality in preschool children in a high malaria transmission setting: community-based, randomised, placebo-controlled trial. *Lancet*. 2006; 367(9505):133–43. PMID: [16413877](https://pubmed.ncbi.nlm.nih.gov/16413877/)
11. Gera T, Sachdev HP. Effect of iron supplementation on incidence of infectious illness in children: systematic review. *Bmj*. 2002; 325(7373):1142. PMID: [12433763](https://pubmed.ncbi.nlm.nih.gov/12433763/)
12. Zimmermann MB, Chassard C, Rohner F, N'Goran E K, Nindjin C, Dostal A, et al. The effects of iron fortification on the gut microbiota in African children: a randomized controlled trial in Cote d'Ivoire. *The American journal of clinical nutrition*. 2010; 92(6):1406–15. doi: [10.3945/ajcn.110.004564](https://doi.org/10.3945/ajcn.110.004564) PMID: [20962160](https://pubmed.ncbi.nlm.nih.gov/20962160/)
13. Black RE, Allen LH, Bhutta ZA, Caulfield LE, de Onis M, Ezzati M, et al. Maternal and child undernutrition: global and regional exposures and health consequences. *Lancet*. 2008; 371(9608):243–60. doi: [10.1016/S0140-6736\(07\)61690-0](https://doi.org/10.1016/S0140-6736(07)61690-0) PMID: [18207566](https://pubmed.ncbi.nlm.nih.gov/18207566/)
14. Laillou A, Prak S, de Groot R, Whitney S, Conkle J, Horton L, et al. Optimal screening of children with acute malnutrition requires a change in current WHO guidelines as MUAC and WHZ identify different patient groups. *PLoS One*. 2014; 9(7):e101159. doi: [10.1371/journal.pone.0101159](https://doi.org/10.1371/journal.pone.0101159) PMID: [24983995](https://pubmed.ncbi.nlm.nih.gov/24983995/)
15. Cambodia Demographic and Health Survey 2010. In: National Institute of Statistics DGfH, and ICF Macro, editor. *Phnom Penh, Cambodia and Calverton, Maryland, USA 2011*.
16. Sinuon M, Anantaphruti MT, Socheat D. Intestinal helminth infections in schoolchildren in Cambodia. *The Southeast Asian journal of tropical medicine and public health*. 2003; 34(2):254–8. PMID: [12971545](https://pubmed.ncbi.nlm.nih.gov/12971545/)
17. Perignon M, Fiorentino M, Kuong K, Burja K, Parker M, Sisokhom S, et al. Stunting, poor iron status and parasite infection are significant risk factors for lower cognitive performance in Cambodian school-aged children. *PLoS One*. 2014; 9(11):e112605. doi: [10.1371/journal.pone.0112605](https://doi.org/10.1371/journal.pone.0112605) PMID: [25405764](https://pubmed.ncbi.nlm.nih.gov/25405764/)
18. *Bench Aids for the diagnosis of intestinal parasites*. Geneva, Switzerland: World Health Organization, 1994.
19. Fagerberg UL, Loof L, Merzoug RD, Hansson LO, Finkel Y. Fecal calprotectin levels in healthy children studied with an improved assay. *Journal of pediatric gastroenterology and nutrition*. 2003; 37(4):468–72. PMID: [14508218](https://pubmed.ncbi.nlm.nih.gov/14508218/)
20. Olsen A, Nawiri J, Friis H. The impact of iron supplementation on reinfection with intestinal helminths and *Schistosoma mansoni* in western Kenya. *Transactions of the Royal Society of Tropical Medicine and Hygiene*. 2000; 94(5):493–9. PMID: [11132373](https://pubmed.ncbi.nlm.nih.gov/11132373/)
21. Stoltzfus RJ, Albonico M, Chwaya HM, Savioli L, Tielsch J, Schulze K, et al. Hemoquant determination of hookworm-related blood loss and its role in iron deficiency in African children. *The American journal of tropical medicine and hygiene*. 1996; 55(4):399–404. PMID: [8916795](https://pubmed.ncbi.nlm.nih.gov/8916795/)
22. Levecke B, Montresor A, Albonico M, Ame SM, Behnke JM, Bethony JM, et al. Assessment of anthelmintic efficacy of mebendazole in school children in six countries where soil-transmitted helminths are endemic. *PLoS neglected tropical diseases*. 2014; 8(10):e3204. doi: [10.1371/journal.pntd.0003204](https://doi.org/10.1371/journal.pntd.0003204) PMID: [25299391](https://pubmed.ncbi.nlm.nih.gov/25299391/)

23. Nikolay B, Brooker SJ, Pullan RL. Sensitivity of diagnostic tests for human soil-transmitted helminth infections: a meta-analysis in the absence of a true gold standard. *International journal for parasitology*. 2014; 44(11):765–74. doi: [10.1016/j.ijpara.2014.05.009](https://doi.org/10.1016/j.ijpara.2014.05.009) PMID: [24992655](https://pubmed.ncbi.nlm.nih.gov/24992655/)
24. Jaeggi T, Kortman GA, Moretti D, Chassard C, Holding P, Dostal A, et al. Iron fortification adversely affects the gut microbiome, increases pathogen abundance and induces intestinal inflammation in Kenyan infants. *Gut*. 2015; 64(5):731–42. doi: [10.1136/gutjnl-2014-307720](https://doi.org/10.1136/gutjnl-2014-307720) PMID: [25143342](https://pubmed.ncbi.nlm.nih.gov/25143342/)
25. Dostal A, Baumgartner J, Riesen N, Chassard C, Smuts CM, Zimmermann MB, et al. Effects of iron supplementation on dominant bacterial groups in the gut, faecal SCFA and gut inflammation: a randomised, placebo-controlled intervention trial in South African children. *The British journal of nutrition*. 2014; 112(4):547–56. doi: [10.1017/S0007114514001160](https://doi.org/10.1017/S0007114514001160) PMID: [24916165](https://pubmed.ncbi.nlm.nih.gov/24916165/)
26. Loukas A, Prociv P. Immune responses in hookworm infections. *Clinical microbiology reviews*. 2001; 14(4):689–703, table of contents. PMID: [11585781](https://pubmed.ncbi.nlm.nih.gov/11585781/)
27. Das JK, Salam RA, Kumar R, Bhutta ZA. Micronutrient fortification of food and its impact on woman and child health: a systematic review. *Systematic reviews*. 2013; 2:67. doi: [10.1186/2046-4053-2-67](https://doi.org/10.1186/2046-4053-2-67) PMID: [23971426](https://pubmed.ncbi.nlm.nih.gov/23971426/)
28. Casey GJ, Montresor A, Cavalli-Sforza LT, Thu H, Phu LB, Tinh TT, et al. Elimination of iron deficiency anemia and soil transmitted helminth infection: evidence from a fifty-four month iron-folic acid and deworming program. *PLoS neglected tropical diseases*. 2013; 7(4):e2146. doi: [10.1371/journal.pntd.0002146](https://doi.org/10.1371/journal.pntd.0002146) PMID: [23593517](https://pubmed.ncbi.nlm.nih.gov/23593517/)
29. Moshage H. Cytokines and the hepatic acute phase response. *The Journal of pathology*. 1997; 181(3):257–66. PMID: [9155709](https://pubmed.ncbi.nlm.nih.gov/9155709/)
30. Weinberg ED. Nutritional immunity. Host's attempt to withhold iron from microbial invaders. *JAMA: the journal of the American Medical Association*. 1975; 231(1):39–41. PMID: [1243565](https://pubmed.ncbi.nlm.nih.gov/1243565/)
31. Cho I, Blaser MJ. The human microbiome: at the interface of health and disease. *Nature reviews Genetics*. 2012; 13(4):260–70. doi: [10.1038/nrg3182](https://doi.org/10.1038/nrg3182) PMID: [22411464](https://pubmed.ncbi.nlm.nih.gov/22411464/)