



Københavns Universitet

Whole-genome sequence of the bacteriophage-sensitive strain *Campylobacter jejuni* NCTC12662

Gencay, Yilmaz Emre; Sørensen, Martine Camilla Holst; Brøndsted, Lone

Published in:
Genome Announcements

DOI:
[10.1128/genomeA.00409-17](https://doi.org/10.1128/genomeA.00409-17)

Publication date:
2017

Document Version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
Gencay, Y. E., Sørensen, M. C. H., & Brøndsted, L. (2017). Whole-genome sequence of the bacteriophage-sensitive strain *Campylobacter jejuni* NCTC12662. DOI: 10.1128/genomeA.00409-17



Whole-Genome Sequence of the Bacteriophage-Sensitive Strain *Campylobacter jejuni* NCTC12662

Yilmaz Emre Gencay, Martine C. H. Sørensen, Lone Brøndsted

Department of Veterinary and Animal Sciences, University of Copenhagen, Frederiksberg, Denmark

ABSTRACT *Campylobacter jejuni* NCTC12662 has been the choice bacteriophage isolation strain due to its susceptibility to *C. jejuni* bacteriophages. This trait makes it a good candidate for studying bacteriophage–host interactions. We report here the whole-genome sequence of NCTC12662, allowing future elucidation of the molecular mechanisms of phage–host interactions in *C. jejuni*.

Campylobacter jejuni is a zoonotic Gram-negative bacterium and the leading cause of foodborne gastroenteritis in the western world. Research within bacteriophages as biocontrols of *C. jejuni* has resulted in the isolation of phages belonging to two genera: *Cp220virus* and *Cp8virus*. *C. jejuni* NCTC12662 is susceptible to most of these phages, including phages dependent on the capsular polysaccharide (CPS) or a motile flagellum for infection (1). Here, we report the genome sequence of NCTC12662 as a common host for *C. jejuni* phages, allowing future molecular investigation and comparison of phage–host interactions in *C. jejuni*.

DNA libraries from the genomic DNA from *C. jejuni* NCTC12662 (obtained from the National Collection of Type Cultures) were prepared using the Nextera XT version 3 kit (Illumina) and sequenced with MiSeq (Illumina) in 2 × 250-bp operating mode. The 550,920 reads generated were *de novo* assembled using CLC Genomics Workbench version 9.5.3, resulting in a total of 47 contigs. Due to the obtained low coverage, another round of sequencing was executed at the Sanger Institute (Cambridge, United Kingdom) using HiSeq 2000 (Illumina), yielding 4,635,024 100-bp raw reads. Subsequently, contigs were joined by the extend and align contig functions using flanking genome data, resulting in an average coverage of 287-fold. The genome sequence was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (2). The circular genome of NCTC12662 is 1,612,586 bp with an average G+C content of 30.7%. It comprises 1,548 coding sequences, 44 tRNA genes, and 3 rRNA operons and carries no prophage-associated genes (3) or plasmids.

Phase-variable expression of genes that carry polyG tracts is a well-known phenomenon for generating phenotypic population diversity in *C. jejuni* (4), and *C. jejuni* NCTC12662 contains 19 polyG tracts. Previously, we found that the phase-variable expression of *cj1421* and *cj1422*, which modify GalNAc and heptose residues of the CPS with *O*-methyl phosphoramidate (MeOPN), respectively, were responsible for phage resistance in *C. jejuni* NCTC11168 (5, 6). NCTC12662 encodes one putative phase-variable MeOPN-transferase (*06810*), showing 83% identity to *Cj1421* and *Cj1422* at the N-terminal part of the protein. Although this indicates that a different CPS residue of NCTC12662 is modified by MeOPN compared to NCTC11168, the phase-variable nature of gene *06810* is conserved in NCTC12662.

NCTC12662 harbors 7 clustered regularly interspaced palindromic repeats (CRISPR) with subtype-II-C CRISPR-associated genes (Cas-1, -2, and -9) sharing high similarity with NCTC11168. However, one of the protospacers is duplicated, and thus NCTC12662 encodes only 5 distinct 31-bp protospacers. Noteworthy, the duplicate protospacer

Received 4 April 2017 Accepted 6 April 2017 Published 25 May 2017

Citation Gencay YE, Sørensen MCH, Brøndsted L. 2017. Whole-genome sequence of the bacteriophage-sensitive strain *Campylobacter jejuni* NCTC12662. Genome Announc 5:e00409-17. <https://doi.org/10.1128/genomeA.00409-17>.

Copyright © 2017 Gencay et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Lone Brøndsted, lobr@sund.ku.dk.

exclusively matches a hypothetical protein (CJE0597) found in *C. jejuni* integrated element 2 (CJIE2) in strain RM1221 (7) and group 4 prophages CJIE4-1 and CJIE4-5 (8). Whether the absence of both groups of prophages in NCTC12662 is due to CRISPR-Cas activity is an intriguing question. Sequence homologies also suggest that NCTC12662 encodes type I, II, III, and IV RM systems (9), indicating no significant effect of these RM systems in *C. jejuni* phage resistance. Thus, further studies are needed to elucidate why NCTC12662 is susceptible to many diverse phages of *C. jejuni*.

Accession number(s). The *C. jejuni* NCTC12662 complete genome is available under GenBank accession no. [CP019965](#)

ACKNOWLEDGMENT

This work was supported by grant 34009-14-0873 (TOPSAFE: Targeted Optimized Phage Solutions for Food Safety) from The Danish AgriFish Agency, Ministry of the Environment and Food, which otherwise had no involvement in the presented work.

REFERENCES

1. Sørensen MCH, Gencay YE, Birk T, Baldvinsson SB, Jäckel C, Hammerl JA, Vegge CS, Neve H, Brøndsted L. 2015. Primary isolation strain determines both phage type and receptors recognised by *Campylobacter jejuni* bacteriophages. *PLoS One* 10:e0116287. <https://doi.org/10.1371/journal.pone.0116287>.
2. Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta)genomic annotation. *Omics* 12:137–141. <https://doi.org/10.1089/omi.2008.0017>.
3. Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. *Nucleic Acids Res* 44:W16–W21. <https://doi.org/10.1093/nar/gkw387>.
4. Bayliss CD, Palmer ME. 2012. Evolution of single sequence repeat-mediated phase variation in bacterial genomes. *Ann N Y Acad Sci* 1267: 39–44. <https://doi.org/10.1111/j.1749-6632.2012.06584.x>.
5. Sørensen MC, van Alphen LB, Harboe A, Li J, Christensen BB, Szymanski CM, Brøndsted L. 2011. Bacteriophage F336 recognizes the capsular phosphoramidate modification of *Campylobacter jejuni* NCTC11168. *J Bacteriol* 193:6742–6749. <https://doi.org/10.1128/JB.05276-11>.
6. Holst Sørensen MC, van Alphen LB, Fodor C, Crowley SM, Christensen BB, Szymanski CM, Brøndsted L. 2012. Phase variable expression of capsular polysaccharide modifications allows *Campylobacter jejuni* to avoid bacteriophage infection in chickens. *Front Cell Infect Microbiol* 2:11. <https://doi.org/10.3389/fcimb.2012.00011>.
7. Fouts DE, Mongodin EF, Mandrell RE, Miller WG, Rasko DA, Ravel J, Brinkac LM, DeBoy RT, Parker CT, Daugherty SC, Dodson RJ, Durkin AS, Madupu R, Sullivan SA, Shetty JU, Ayodeji MA, Shvartsbeyn A, Schatz MC, Badger JH, Fraser CM, Nelson KE. 2005. Major structural differences and novel potential virulence mechanisms from the genomes of multiple *Campylobacter* species. *PLoS Biol* 3:e15. <https://doi.org/10.1371/journal.pbio.0030015>.
8. Clark CG, Chong PM, McCorrister SJ, Mabon P, Walker M, Westmacott GR. 2014. DNA sequence heterogeneity of *Campylobacter jejuni* CJIE4 prophages and expression of prophage genes. *PLoS One* 9:e95349. <https://doi.org/10.1371/journal.pone.0095349>.
9. Gardner SP, Olson JW. 2012. Barriers to horizontal gene transfer in *Campylobacter jejuni*, p 19–42. In Gadd G, Sariaslani S (ed), *Advances in applied microbiology*, 1st ed. Academic Press, San Diego, CA.