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Neochloris oleoabundans cell walls have an altered composition when cultivated under different growing conditions

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ABSTRACT

The impact that environmental factors have on the intracellular components of microalgae has been the focus of research for a number of decades. Despite that, their effects on the cell wall have received very little attention. In this study, we investigated how different growing conditions affect the cell walls of N. oleoabundans. The results revealed that the cell wall composition varied in that the modifications were different in the four cultivation media: freshwater nitrogen-replete (optimum culture) and -depleted conditions, and seawater nitrogen-replete and -depleted conditions. Nitrogen deficiency in freshwater cultivation was the only condition that significantly (p < .05) increased the total content of carbohydrates in the cell wall. The three most abundant components of freshwater-cultivated cell wall polysaccharides were rhamnose, galactose and glucuronic acid whereas in seawater media the main components of cell wall polysaccharides were rhamnose, glucose and galactose. The combined results of the biochemical analyses and monoclonal antibodies epitope-binding revealed that N. oleoabundans cell walls are likely composed of sulphated polysaccharides enriched in mannose, β-(1→4)-α-mannans, and glucose as they grow in seawater. Salinity and nitrogen deficiency also had an impact on the nitrogenous components of the cell wall. Under these conditions we observed a decrease in glucosamine in the cell wall. The analysis of specific binding of monoclonal antibodies, revealed that the cell wall of N. oleoabundans is possibly enriched in arabinogalactan proteins (AGPs). Under salinity and nitrogen deficiency N. oleoabundans increased the proportion of the non-polar to polar amino acids in the cell walls. An increase of leucine in the cell walls may suggest that N. oleoabundans contains leucine-rich repeat proteins which are known to play a vital role in stress responses. This report provides new insights into microalgae cell wall biology and how cell walls are remodelled when growing under different conditions.

1. Introduction

The impact that environmental factors have had on the intracellular components of microalgae has already been the subject of research for a number of decades. In contrast, variations in cell wall composition and structure due to the impact of different environments have received limited attention. Adverse environmental factors can influence physiological processes and consequently have an impact on the regulation of cell wall biosynthesis, at both transcriptional and biochemical levels [1,2].

Nitrogen deficiency and salinity are two stress factors that have been used to modulate the intracellular composition in various microalgae species [3–7]. It is clear that these factors play a significant role in the cell carbon partitioning, and one would expect these alterations to be visible in the cell wall as well. A few reports are now available on the effects of stress conditions on the algae cell wall. These include alterations in the cell wall composition or structure such as increased cell wall thickness which is associated with nitrogen deficiency [8,9]. A great example on effects of stress condition on green microalgae cell wall has been recently published [2]. It was revealed that the increased thickness of the cell wall caused by nitrogen deficiency in Nannochloropsis salina is associated with up-regulation of genes encoding for cellulose biosynthetic enzymes, which resulted in an increase in the cellulose content of the cell wall. Another study demonstrated that transition of N. salina from high to low saline culture increased the cell wall thickness, although detailed changes in biochemical composition are not yet available [10].

Neochloris oleoabundans, is oleaginous unicellular green microalgae belonging to the Chlorophyta phylum. Over the years, this microalgae has been considered one of the most promising candidate industrial
microalgae, due its high growth rate and biomass composition [11]. *N. oleoabundans* is an epiphytic freshwater green microalgae which was originally isolated from the sand dunes in Saudi Arabia, an environment where the lack of water is a tenacious threat and cells can be exposed to saline or drought stress [12]. The cell wall is the outermost structure of the cell and the first part of the cell to be exposed to severe conditions. It therefore requires specific properties to guarantee cell viability. Owing to the plasticity of *N. oleoabundans* and its saline resistance mechanism, this alga can be cultivated in both freshwater and saline cultivation media, with seawater salt concentration [13–15].

Recently, the cell wall composition of *N. oleoabundans* growing in freshwater condition was biochemically characterized and the morphology of the cell wall dissected by means of electron microscopy [16]. This report revealed that 56% of the cell wall is composed of carbohydrates and nitrogenous components such as amino sugars and proteins.

In this manuscript we aim to evaluate the effects of different cultivation media, freshwater nitrogen-replete (optimum culture) and -depleted conditions, and seawater nitrogen-replete and -depleted, on cell wall composition, with the main focus on carbohydrate and protein compositions. We expect that a lack of nitrogen could not only affect the carbohydrates but also have a direct impact on the nitrogenous components of the cell wall, such as amino sugars and proteins. The results revealed considerable variations in the biochemical composition of cell walls grown under different culture conditions. These results provide important insights into the biology of *N. oleoabundans* and will greatly contribute to our understanding of cell wall remodelling in green microalgae when they are exposed to adverse environments.

### 2. Materials and methods

#### 2.1. Biomass supply

*N. oleoabundans* (UTEX 1185, University of Texas Culture Collection of Algae) was pre-cultured in 100 mL sterilized fresh or seawater medium. Subsequent to reaching an optimum density (~0.2 g/L), the culture was used to inoculate a vertical tubular photobioreactor (VT LGem, 1300 L working volume, Rotterdam, The Netherlands) installed inside a greenhouse at AlgaePARC facilities (Wageningen, The Netherlands). Details of the reactor are available on the manufacturer's website (www.lgem.nl). The same inoculations were used for the nitrogen-depleted experiments. The reactor was operated in a batch phase with the use of artificial light (7.2 kW), at an average temperature of 25 °C and pH of 7.5. Concentration of the nutrients in the medium can be found in Table 1. The culture was monitored daily and the biological parameters including dry matter, optical density (530, 680, 750 nm) and quantum yield of the harvested points were recorded (Supplementary file 1). Additionally, daily microscopic visualization in order to assess the cell morphology and possible contamination was conducted. Details of the measurements have already been described [16]. Biomass from different points in time of this batch pipeline production was harvested, centrifuged (80 Hz, ~3000g, 0.75 m/s/1) using a spiral plate centrifuge (Evodos 10, Raamsdonksveer, The Netherlands), rinsed with water and dried in an oven at 60 °C until a constant weight was achieved. The dried biomass at different points of harvesting was pooled and used as a starting material. Cell wall extraction was carried out in three replicates. The extracted cell wall materials were combined and used for further biochemical characterization. All the biochemical analyses mentioned in this publication were performed at least with two technical replications and the values presented are the mean ± standard deviation (SD).

#### 2.2. Preparing samples to extract the cell wall

Cell wall preparation was carried out as described in [16]. In brief, 1 g of dried biomass was mechanically disrupted in a mill for 1 min at a frequency of 25 s−1 (Mixer Mill MM 200-Retsch, Germany). Following this, three incubation cycles of chloroform: methanol (2:1) was used in order to remove the intracellular lipids. Each cycle was conducted at 60 °C for 30 min continuously shaking at 600 rpm. After each incubation, samples were centrifuged and the supernatant was discarded. Subsequent to the last extraction, the residual pellets were dried in an oven at 60 °C until a constant weight was achieved. When the lipids had been removed from the sample, the biomass was de-starched by incubation in a buffer containing a cocktail of alpha-amylase. In summary, 25 mL of maleate buffer (0.01 M C4H4O4, 0.01 M NaCl, 0.001 M CaCl2, and 0.05% w/v NaN3) at pH 6.5 was added to the sample and then incubated for 90 min at 85 °C. Once the sample had cooled down to room temperature, a cocktail of alpha-amylase (50 μL/25 mL, ANKOM Technology Corporation, Fairpoint, NY) was added to the suspension which was then incubated for 24 h at 30 °C. Subsequently, samples were centrifuged and the supernatant containing glucose derived from starch was discarded. All the centrifugal steps mentioned in this publication were conducted at 4200g for 10 min unless stated otherwise.

#### 2.3. Neutral Detergent Fibre (NDF) extraction of the cell wall

Cells walls from the oil-free de-starched sample were extracted in accordance with the established protocol developed by Ankom Technology (ANKOM Technology Corporation, Fairpoint, NY) as described in [16]. In short, the biomass was incubated at 100 °C for 1 h in a 25 mL NDF buffer (104 mM sodium dodecyl sulphate, 50 mM ethylene-diaminetetraacetic disodium salt (dihydrate), 17.8 mM sodium borate, 32 mM sodium phosphate dibasic (anhydrous), 79 mM sodium sulphite and 10 g/L triethyleneglycol) and stirred constantly at 600 rpm. As soon as the incubation was accomplished, the suspension was then centrifuged and the supernatant discarded. The remaining pellets were washed twice using ultrapure water (Milli-Q®) and once with ethanol (96%). Subsequently, the pellets were then dried in an oven at 60 °C until a constant weight was achieved. These pellets corresponded to the total cell wall and will be further referred as the NDF-cell wall.

### Table 1

<table>
<thead>
<tr>
<th>Component</th>
<th>Freshwater</th>
<th>Artificial seawater</th>
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</thead>
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<tr>
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<td></td>
</tr>
<tr>
<td>NaNO3</td>
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<td>11.765</td>
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<td>NaCl</td>
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<td>419.233</td>
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<tr>
<td>CaCl2·2H2O</td>
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<td></td>
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<tr>
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<td></td>
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<tr>
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<tr>
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<tr>
<td>KI</td>
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</table>
2.4. Sulphuric acid hydrolysis and characterization of the cell wall carbohydrates

A total amount of carbohydrates in the NDF-cell wall of N. oleoabundans were measured subsequent to the sulphuric acid hydrolysis of polysaccharides as described earlier [16]. Briefly, 20 mg of dried NDF-cell wall was incubated in 1 mL of 72% (V/V) H2SO4 for 1 h at 30 °C. The acid concentration was then diluted by adding the right amount of ultrapure water (Milli-Q) to reach the final concentration of 6% (V/V). Incubation with the diluted acid concentration was further continued for 1 h at 121 °C (autoclave). Following the hydrolysis, the hydrolysates were cooled down, neutralized and after filtration using a 0.45 μm filter, sugar content was analysed by means of High Performance Anion Exchange Chromatography (HPAEC, Dionex ICS5000 + DC, CarboPac PA1, 2 × 250 mm, Thermo Fisher Scientific, Waltham, MA, USA) as described in [16].

2.5. Immunolabelling of the N. oleoabundans cell wall components

Comprehensive microarray polymer profiling (CoMPP) of N. oleoabundans grown in either fresh or seawater nitrogen-replete was performed as previously described in [17–19]. Cell wall was isolated using a alcohol insoluble residue (AIR) procedure and used as a starting materials to extract the carbohydrates microarrays [20,21]. In brief, the cell wall polymers were extracted sequentially from 10 mg of the AIR-cell wall using 50 mM 1,2-diaminocyclohexanetetraacetic acid (CDTA), pH 7.5, followed by extraction with 4M NaOH with 0.1% m/V NaBH4, and extractions printed in four dilutions and two replicates giving a total of 8 spots per sample. The same amount of cell wall material was used for each sample. Nitrocellulose microarrays were printed as described previously [17,19]. Briefly, the printed arrays were probed with a panel of anti-rat and anti-mouse monoclonal antibodies (PlantProbes and Biosupplies). Antibodies were diluted in PBS containing 5% w/v milk powder to 1/10 and 1/1000, respectively. For secondary antibodies, anti-rat and anti-mouse secondary antibodies conjugated to alkaline phosphatase (Sigma) were diluted in MPBS to 1/5000. Developed microarrays were scanned at 2400 dpi (CanoScan 8800F), converted into TIFFs and signals were measured using Array-Pro Analyzer 6.3 (Media Cybernetics). Data are shown in a heatmap in which the colour intensity is correlated to a mean spot signal value. A cut off, of 5 arbitrary units was used.

2.6. Proteins and inorganic components of the cell wall

Potential changes in the nitrogenous components of the cell wall due to the deficiency of nitrogen in the culture were further explored. To this end, the protein content was measured as described in [16]. In summary, a sample of 150 mg NDF-cell wall was dried in a convection oven for 1 h at 100 °C and after cooling it down in a desiccator, the total nitrogen content was determined by combustion at 950 °C using a LECO analyser (LECO CN 628 Dumas analyser, LECO Corporation, USA). Total protein content of the cell wall was assessed by multiplying the Nitrogen to Protein (NTP) conversion factor with the nitrogen content. This conversion factor was calculated specifically for the NDF-cell wall of N. oleoabundans as reported previously [16]. Prior to characterizing the amino acids, a 20 mg sample of the NDF-cell wall was hydrolysed in 6 N HCl for 24 h at 100 °C. Subsequent to hydrolysis, the amino acid characterization was carried out using the Gas Chromatography (GC) technique based on the EZfaast™ method (Phenomenex Inc.). All the steps were carried out in line with the protocol annexed to the kit [22].

Possible changes in the inorganic material of the cell wall under different environments were investigated. The inorganic content was assessed in line with the protocol described in [16]. Briefly, 100 mg ground dried NDF-cell wall in a pre-weighted glass tube was heated gradually in a muffle furnace for 5 h at 575 °C and the contents were turned into ash. The inorganic residue was then cooled down to room temperature in a desiccator and the weight was recorded. To measure the cations and anions, approximately 3 mg of ash was dissolved in 1 mL of 3 M formic acid and incubated for 15 min at 99 °C. Subsequent to dissolving, the solution was diluted 10 times with MQ water, filtered using a 0.45 μm filter and then the ions were characterized using the Ion Chromatography (IC) system 850 Professional (Metrohm Switzerland). The anions were determined by means of a Metrosep A 150, 150/4.0 mm column equipped with a Metrosep C5/5 Supp 4/6 Guard column and the cations with a Metrosep C4 Supp 4, 250/4.0 mm column equipped with a Metrosep A Supp 4/6 Guard column.

2.7. Statistical analysis

Significant difference between the each trait under the different growing conditions was assessed by two-way analysis of variance (ANOVA). The Fisher’s unprotected least significant difference values were calculated at 5% probability. All statistical analyses were performed using Genstat (19th edition software, VSN International, Hemel Hempstead, UK).

3. Results

3.1. Composition of the cell wall carbohydrates alters according to different growing conditions

Analysis of the N. oleoabundans cell cultivated in the 4 different media resulted in significant differences (p < .05) in the total content of cell walls between the growing conditions evaluated (Fig. 1). Nevertheless, the total amount of carbohydrates in the cell wall did not show any significant differences (p < .05) between the different treatments with the exception of cell cultivated in fresh water with nitrogen deficiency (Fig. 2). Under this condition, the total carbohydrates in the cell wall were about 14% (percentage of NDF-cell wall). Rhamnose, arabinose, glucosamine, galactose, glucose, mannose, xylose and glucuronic acid were the monosaccharides detected in the cell wall of N. oleoabundans independently of the media used for cultivation (Fig. 3 and Supplementary file 2). However, the proportion of the different monosaccharides was different. Rhamnose, galactose and glucuronic acid were the three most abundant monosaccharides in freshwater cultivations (~68% of total monosaccharides), whereas in seawater cultivations rhamnose, glucose and galactose constituted the main cell wall monosaccharides (65% of total monosaccharides). The proportion of monosaccharides remained relatively constant when cells were cultivated in seawater conditions (Nitrogen-deplete versus nitrogen-replete). Nitrogen depletion in freshwater cultivations resulted
in a significant increase in the amount of arabinose and glucuronic acid and a reduced galactose content (p < .05). Except for xylose, of which the content was similar in all growing conditions, we observed that there were alterations in the relative amount of almost all monosaccharides in seawater cultivations, with or without nitrogen, as compared to the corresponding samples cultivated in freshwater conditions. Our results disclosed that glucose and mannose are the monosaccharides which increased the most in the cell wall carbohydrates of cells cultivated in saline media (p < .05).

CDTA and NaOH-extracted glycans of N. oleoabundans cell wall, fresh and seawater nitrogen-replete cultivations, were probed with 38 monoclonal antibodies (mAbs). Fig. 4 shows a heatmap of the relative abundance of the mAbs binding to specific cell wall components. Overall, the majority of the mAbs did not bind to the N. oleoabundans cell wall polymers. Walls of cell cultivated in both nitrogen-replete freshwater and seawater conditions revealed that they were composed of (1→4)-β-D-xylan/arabinoxylan. Additionally, our results indicated the possible existence of arabinogalactan proteins (AGPs) in the cell wall of N. oleoabundans. Monoclonal antibodies of JIM16 and LM14 showed a high binding affinity with the NaOH-extract of the freshwater cell wall (nitrogen-replete), whereas JIM13 indicated a high affinity with the CDTA-extract of seawater cell wall (nitrogen-replete). Remarkably, our results demonstrated the accumulation of (1→4)-β-D-

mannan/galactomannan in N. oleoabundans cell walls grown in seawater condition, whereas these epitopes were either absent or in-accessible in freshwater cultivation.

3.2. Cell walls have a varied protein composition when grown under different conditions

Analysis of the protein content in the cell walls of microalgae cultivated under different conditions revealed that cell wall accumulated a high amount of proteins in both sea and freshwater cultivation provided there was a sufficient amount of nitrogen in the medium (Fig. 2 and Supplementary file 3). However, under nitrogen-depleted growing conditions, the protein content of the cell wall decreased significantly (p < .05), in which nitrogen-depleted seawater showed the lowest content (19.3% of the NDF-cell wall). Further amino acid characterization revealed a decrease in the algae cell wall polar amino acids that were cultivated in fresh water under nitrogen depleted condition (Fig. 5). This reduction was specifically on the positive-side-chain amino acids, where lysine decreased dramatically and histidine reduced to undetectable amounts, as well as the polar-uncharged amino acids where the larger reductions were observed in serine and threonine.

Cells cultivated in nitrogen-depleted seawater revealed a higher content of polar amino acids in the cell wall relatively to the nitrogen-replete culture (p < .05). Positive-side-chain-amino acids were not detected in the cell wall of a nitrogen-depleted seawater cultivation. Aspartic acid and isoleucine remained constant in the different growing conditions.

Results of the inorganic portion of the cell wall revealed a higher accumulation of inorganic components in the nitrogen deficient cultures (Fig. 2). Under nitrogen-replete growing conditions, the inorganic content of the cell wall decreased, with the lowest percentage being found in freshwater cultivation (~5%). Ion chromatography analysis revealed that the cell wall is primarily composed of sulphate and sodium, adding up to almost 85% of the total ash content in a nitrogen-replete freshwater culture and 98% in other growing conditions (Fig. 6). Phosphate and magnesium were only detected in nitrogen-replete freshwater culture.

4. Discussion

Adverse growing conditions such as salinity and nitrogen depletion can influence cell physiology and consequently cell wall composition [1]. We observed significant variation in the total content of cell walls between the growing conditions assessed (4%–19% g/g, p < .05). Under different growing conditions N. oleoabundans cells were able to
change the composition of their cell walls. This variation highlights an important property of N. oleoabundans cell walls, which are plastic and can adapt to different growing conditions. In this study we have characterized the content and composition of carbohydrates, proteins and inorganic components, though microalgae cell wall might have other components, such as lipid, that may as well vary under the adverse growing conditions, but these were not object of our study.

Our results revealed that walls of N. oleoabundans cells cultivated under nitrogen-depleted conditions accumulated a higher content of carbohydrates, although this increase was not significant (p < .05) in the seawater cultivation (Fig. 2). Previous studies of other microalgae indicate that when cells were cultivated under nitrogen-depleted conditions an increase in polysaccharide content of the cell wall was observed [2,23,24]. These results illustrate the ability of the N. oleoabundans cell wall to function as an additional type of sink for photoassimilates accumulated under nitrogen stress. Seawater cultivation resulted in alterations in the monosaccharide profile of the cell wall as compared to the freshwater culture (Fig. 3). Glucose and mannose content in the cell wall were abundantly increased when N. oleoabundans was cultivated in seawater (p < .05). We have observed that the majority of the mAbs did not bind to the N. oleoabundans cell wall polymers. The most probable explanation is the absent of the epitopes...
in N. oleoabundans cell wall, which would not be surprising as these antibodies have been developed for cell walls of plants and other algae species. Another explanation could be that the epitopes might have been inaccessible and/or possibly degraded during the extraction procedure. Nevertheless, results from monoclonal antibodies disclosed that \((1 \rightarrow 4)\)-\(\beta\)-\(\alpha\)-mannan/galactomannan are the possible epitopes of the accumulated mannan in the seawater cultivated cell wall. Although results from monosaccharides composition of the cell wall revealed the presence of mannose in a freshwater cultivation (~7% of total monosaccharides, g/g), evaluated monoclonal antibodies were unable to recognize the mannose-epitope(s). This may be attributable to the existence of different epitope of mannose or inaccessibility of the same mannose-epitope in the cell wall of freshwater cultivation. Previous studies on marine algae established that some microalgae are enriched in sulphated polysaccharides including \(\beta\)-(1 \rightarrow 4)-\(\alpha\)-mannans, \((1 \rightarrow 3)\)-\(\beta\)-\(\alpha\)-arabinopyranans and other sulphated polysaccharides containing galactose, glucose and arabinose [25-28]. Marine algae are able to resist the saline environment on account of particular mechanisms such as sodium exclusion or accumulation of sulphated polysaccharides [27-30]. The latter mechanism, which is unique to marine algae, is considered to be a strategy of adaptation in marine territories and is also present in (some) halophyte terrestrial plants [27,28]. It has been reported that sulphated polysaccharides, which provide a gel-like matrix for a fibrous and crystalline component of the cell wall, are part of the cell walls of some marine species belonging to the phylum Chlorophyta [28,31]. Considering the abundance of mannose, \(\beta\)-(1 \rightarrow 4)-\(\alpha\)-mannans, and glucose together with a high content of inorganic sulphate in the seawater cultivated N. oleoabundans cell wall, it is tempting to hypothesize the presence of sulphated polysaccharides which may contribute to the adaptation of this species in a saline medium.

Nitrogen-containing biopolymers are one of the main components in the microalgae cell wall belonging to the Chlorococcaleae, therefore it is not surprising that a restricted amount of nitrogen in the medium caused a reduction of the cell wall fraction in the total cell mass of N. oleoabundans [32-34]. As depicted in Fig. 3 there is a considerable reduction of glucosamine in the cell walls when they are cultivated under nitrogen depletion or saline medium. A recent transcriptomic study of N. oleoabundans cultivated under nitrogen depletion indicated a downregulation of glucosamine fructose-6-phosphate aminotransferase (GFA) [35]. This enzyme is involved in the hexosamine (amino sugar) biosynthesis pathway, synthesizing glucosamine-6-phosphate (GlcN-6P) from fructose-6-phosphate (Fru-6P) which is derived from glucose. It has been demonstrated that nitrogen depletion in Chlorella vulgaris resulted in morphological alteration of the cell wall [36]. C. vulgaris growing in nitrogen-replete conditions contains hair-like fibres mounted on the outer layer of the cell wall, referred to as hyaluron, an unsulphated glycosaminoglycan structure composed of \(\beta\)-1,4-glucuronic acid and \(\beta\)-1,3-N-acetylglucosamine [36-38]. Under nitrogen-depleted conditions C. vulgaris lacks these hair-like fibres.

The change in cell wall mass under nitrogen-depleted conditions was predominantly attributed to a reduction in the protein content of the cell wall (Figs. 1 and 2). Localized cell wall proteins comprise enzymes, both in situ enzymes for development and remodelling, and enzymes involved in biotic and abiotic responses; as well as glycine-rich proteins (GRPs), proline-rich proteins, extensins and hydroxyproline-rich glycoproteins (HRGs) [39,40]. Arabinogalactan proteins (AGPs) belong to the HRGs and are found at the cell surface of a wide variety of plants and algae [34,40,41]. The positive signal of the antibodies JIM16, JIM13 and JIM14 toward the arabinogalactan/arabinogalactan-protein antigens revealed the possible existence of AGPs in N. oleoabundans cell walls. In line with previous studies, these monoclonal antibodies have an affinity to AGP1 and AGP2 immunogens [42,43]. The likely existence of arabinogalactan proteins in the N. oleoabundans cell wall supports its taxonomical classification into the phylum Chlorophyta, in which cell walls are known to contain AGPs [44]. Despite these enlightening findings, questions still remain concerning the particular biological function of AGP1 and AGP2 in the N. oleoabundans cell wall. The exact function of AGPs in plant cells is still a matter of debate, yet cell division, cell extension, abiotic stress tolerance and cell viability are the main functions discussed in literature [41,45,46].
Polar amino acids were more abundant in the walls of cells cultivated in nitrogen-replete freshwater in comparison to all the other growing conditions (Fig. 5). The final structure of a protein comprises polar/hydrophilic side-chain amino acids, which are normally located on the surface of the proteins and are in contact with the aquatic milieu, and non-polar/hydrophobic side-chain amino acids existing on the interface [47]. Changes in the abundance of amino acids, each with the different chemistry of side chain, alters conformation and structure of the protein. Amongst non-polar amino acids, the increase of leucine was considerable in the cell wall of N. oleoabundans grown in either seawater conditions or freshwater nitrogen-depleted culture. The significant increase of this amino acid (p < .05) could indicate that N. oleoabundans cell walls contain leucine-rich repeat proteins. These proteins are part of a large variety of organisms and are reported to be involved in many developmental processes and responses to biotic and abiotic stresses [48–50]. Due to the intrinsic limitation of the EZFaaSTM method, we were unable to detect asparagine, glutamine and arginine in the N. oleoabundans cell wall. Furthermore, our results revealed a lack of cysteine and tryptophan in the N. oleoabundans cell wall that may well have been degraded during the acid hydrolysis process.

Our findings herein clearly confirm that the cell wall composition of N. oleoabundans varies depending on the cultivation medium. Remodelling of this dynamic structure is key for the plasticity of this species to survive in a wide range of growing conditions.

5. Conclusion

In this study we have highlighted the importance of the N. oleoabundans cell wall in response to high saline and/or nitrogen-deficient mediums. Cell wall remodelling under saline conditions comprises the possible accumulation of sulphated polysaccharides enriched in mannos, β-(1→4)-α-mannans, and glucose. The likely abundance of sulphated polysaccharides together with non-polar amino acids, especially leucine, could well enable the cell to resist the saline environment. Nitrogen depletion also has an important effect on the cell wall composition, being the primary effect of this stress a substantial reduction in the nitrogenous components of the cell wall. This is the first study of this kind on the cell wall biology of N. oleoabundans which has enabled us to understand the complexity of remodelling the cell wall in response to salt and/or nitrogen deficiency.

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Author contributions

B.R performed the experiments, analysed data and wrote the paper. L.M.T wrote and coordinated the project, supervised the research and helped shaping the manuscript. A.D helped developing protocols and assisted in the chemical analysis. M.G.R and B.J performed the carbohydrate microarray and the glycomic profile and revised the manuscript. All authors read and approved the final manuscript.

Conflict of interest

The authors declare that there is no conflict of interest for this paper.

Informed consent

The authors declare that there are no conflicts, informed consent, human or animal rights applicable for this paper.

Authorship consent

All authors read, approved the final manuscript and agreed for its submission for peer review.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.algal.2019.101482.

References


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